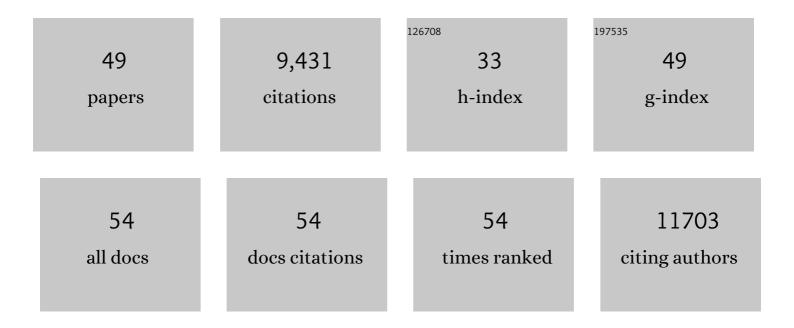
Jeremy E Wilusz

List of Publications by Year in descending order

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IFDEMV F WILLISZ

| # | Article | IF | CITATIONS |
|----|--|------|-----------|
| 1 | Long noncoding RNAs: functional surprises from the RNA world. Genes and Development, 2009, 23, 1494-1504. | 2.7 | 2,032 |
| 2 | Short intronic repeat sequences facilitate circular RNA production. Genes and Development, 2014, 28, 2233-2247. | 2.7 | 773 |
| 3 | 3′ End Processing of a Long Nuclear-Retained Noncoding RNA Yields a tRNA-like Cytoplasmic RNA. Cell, 2008, 135, 919-932. | 13.5 | 597 |
| 4 | <i>MEN ε/β</i> nuclear-retained non-coding RNAs are up-regulated upon muscle differentiation and are essential components of paraspeckles. Genome Research, 2009, 19, 347-359. | 2.4 | 570 |
| 5 | A Circuitous Route to Noncoding RNA. Science, 2013, 340, 440-441. | 6.0 | 458 |
| 6 | Combinatorial control of <i>Drosophila</i> circular RNA expression by intronic repeats, hnRNPs, and SR proteins. Genes and Development, 2015, 29, 2168-2182. | 2.7 | 419 |
| 7 | A triple helix stabilizes the 3′ ends of long noncoding RNAs that lack poly(A) tails. Genes and Development, 2012, 26, 2392-2407. | 2.7 | 375 |
| 8 | A 360° view of circular RNAs: From biogenesis to functions. Wiley Interdisciplinary Reviews RNA, 2018, 9, e1478. | 3.2 | 356 |
| 9 | Sensing Self and Foreign Circular RNAs by Intron Identity. Molecular Cell, 2017, 67, 228-238.e5. | 4.5 | 346 |
| 10 | Non-AUG translation: a new start for protein synthesis in eukaryotes. Genes and Development, 2017, 31, 1717-1731. | 2.7 | 322 |
| 11 | The Output of Protein-Coding Genes Shifts to Circular RNAs When the Pre-mRNA Processing Machinery Is Limiting. Molecular Cell, 2017, 68, 940-954.e3. | 4.5 | 319 |
| 12 | Circular RNA CircFndc3b modulates cardiac repair after myocardial infarction via FUS/VEGF-A axis. Nature Communications, 2019, 10, 4317. | 5.8 | 280 |
| 13 | A length-dependent evolutionarily conserved pathway controls nuclear export of circular RNAs. Genes and Development, 2018, 32, 639-644. | 2.7 | 238 |
| 14 | High-Resolution Mapping of RNA-Binding Regions in the Nuclear Proteome of Embryonic Stem Cells. Molecular Cell, 2016, 64, 416-430. | 4.5 | 226 |
| 15 | Biogenesis and Functions of Circular RNAs Come into Focus. Trends in Cell Biology, 2020, 30, 226-240. | 3.6 | 224 |
| 16 | Long noncoding RNAs: Re-writing dogmas of RNA processing and stability. Biochimica Et Biophysica Acta - Gene Regulatory Mechanisms, 2016, 1859, 128-138. | 0.9 | 182 |
| 17 | The Integrator Complex Attenuates Promoter-Proximal Transcription at Protein-Coding Genes. Molecular Cell, 2019, 76, 738-752.e7. | 4.5 | 150 |
| 18 | tRNAs Marked with CCACCA Are Targeted for Degradation. Science, 2011, 334, 817-821. | 6.0 | 139 |

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| # | Article | IF | CITATIONS |
|----|---|------|-----------|
| 19 | An improved method for circular RNA purification using RNase R that efficiently removes linear RNAs containing G-quadruplexes or structured 3′ ends. Nucleic Acids Research, 2019, 47, 8755-8769. | 6.5 | 130 |
| 20 | A 3′ Poly(A) Tract Is Required for LINE-1 Retrotransposition. Molecular Cell, 2015, 60, 728-741. | 4.5 | 120 |
| 21 | The Integrator complex cleaves nascent mRNAs to attenuate transcription. Genes and Development, 2019, 33, 1525-1538. | 2.7 | 113 |
| 22 | Circular RNAs: Unexpected outputs of many protein-coding genes. RNA Biology, 2017, 14, 1007-1017. | 1.5 | 111 |
| 23 | Tissue-Dependent Expression and Translation of Circular RNAs with Recombinant AAV Vectors InÂVivo. Molecular Therapy - Nucleic Acids, 2018, 13, 89-98. | 2.3 | 89 |
| 24 | Biogenesis and Regulatory Roles of Circular RNAs. Annual Review of Cell and Developmental Biology, 2022, 38, 263-289. | 4.0 | 75 |
| 25 | CRISPR/Cas13 effectors have differing extents of off-target effects that limit their utility in eukaryotic cells. Nucleic Acids Research, 2022, 50, e65-e65. | 6.5 | 63 |
| 26 | An Unchartered Journey for Ribosomes: Circumnavigating Circular RNAs to Produce Proteins. Molecular Cell, 2017, 66, 1-2. | 4.5 | 62 |
| 27 | On-Enzyme Refolding Permits Small RNA and tRNA Surveillance by the CCA-Adding Enzyme. Cell, 2015, 160, 644-658. | 13.5 | 61 |
| 28 | Ribosome queuing enables non-AUG translation to be resistant to multiple protein synthesis inhibitors. Genes and Development, 2019, 33, 871-885. | 2.7 | 60 |
| 29 | Controlling translation via modulation of <scp>tRNA</scp> levels. Wiley Interdisciplinary Reviews RNA, 2015, 6, 453-470. | 3.2 | 59 |
| 30 | An unexpected ending: Noncanonical $3\hat{a}\in^2$ end processing mechanisms. Rna, 2010, 16, 259-266. | 1.6 | 54 |
| 31 | Repetitive elements regulate circular RNA biogenesis. Mobile Genetic Elements, 2015, 5, 39-45. | 1.8 | 54 |
| 32 | A conserved virus-induced cytoplasmic TRAMP-like complex recruits the exosome to target viral RNA for degradation. Genes and Development, 2016, 30, 1658-1670. | 2.7 | 49 |
| 33 | TET2 chemically modifies tRNAs and regulates tRNA fragment levels. Nature Structural and Molecular Biology, 2021, 28, 62-70. | 3.6 | 42 |
| 34 | Best practices to ensure robust investigation of circular RNAs: pitfalls and tips. EMBO Reports, 2021, 22, e52072. | 2.0 | 37 |
| 35 | Engineering highly efficient backsplicing and translation of synthetic circRNAs. Molecular Therapy - Nucleic Acids, 2021, 23, 821-834. | 2.3 | 36 |
| 36 | The Integrator Complex in Transcription and Development. Trends in Biochemical Sciences, 2020, 45, 923-934. | 3.7 | 35 |

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| # | Article | IF | CITATIONS |
|----|---|------|-----------|
| 37 | Inducible Expression of Eukaryotic Circular RNAs from Plasmids. Methods in Molecular Biology, 2017, 1648, 143-154. | 0.4 | 30 |
| 38 | Removing roadblocks to deep sequencing of modified RNAs. Nature Methods, 2015, 12, 821-822. | 9.0 | 24 |
| 39 | Chimeric peptide nucleic acid compounds modulate splicing of the bcl-x gene in vitro and in vivo. Nucleic Acids Research, 2005, 33, 6547-6554. | 6.5 | 23 |
| 40 | The Negative Regulator of Splicing Element of Rous Sarcoma Virus Promotes Polyadenylation. Journal of Virology, 2006, 80, 9634-9640. | 1.5 | 23 |
| 41 | The capping enzyme facilitates promoter escape and assembly of a follow-on preinitiation complex for reinitiation. Proceedings of the National Academy of Sciences of the United States of America, 2019, 116, 22573-22582. | 3.3 | 16 |
| 42 | Circle the Wagons: Circular RNAs Control Innate Immunity. Cell, 2019, 177, 797-799. | 13.5 | 14 |
| 43 | Noncoding RNAs: biology and applications—a Keystone Symposia report. Annals of the New York Academy of Sciences, 2021, 1506, 118-141. | 1.8 | 13 |
| 44 | RNAi Screening to Identify Factors That Control Circular RNA Localization. Methods in Molecular Biology, 2021, 2209, 321-332. | 0.4 | 5 |
| 45 | Ending on a high note: Downstream ORFs enhance mRNA translational output. EMBO Journal, 2020, 39, e105959. | 3.5 | 5 |
| 46 | Nonsense-mediated RNA decay: at the †̃cutting edge' of regulated snoRNA production: Figure 1 Genes and Development, 2014, 28, 2447-2449. | 2.7 | 4 |
| 47 | Attenuation of Eukaryotic Protein-Coding Gene Expression via Premature Transcription Termination. Cold Spring Harbor Symposia on Quantitative Biology, 2019, 84, 83-93. | 2.0 | 4 |
| 48 | Use of circular RNAs as markers of readthrough transcription to identify factors regulating cleavage/polyadenylation events. Methods, 2021, 196, 121-128. | 1.9 | 2 |
| 49 | A04â€Circhtt, a circular rna from the huntington's disease gene locus: functional characterization and possible implications for disease modulation. , 2021, , . | | 0 |