## Fedor V Subach

List of Publications by Year in descending order

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FEDOR V SUBACH

#	Article	IF	CITATIONS
1	Photoactivatable mCherry for high-resolution two-color fluorescence microscopy. Nature Methods, 2009, 6, 153-159.	9.0	569
2	Conversion of Red Fluorescent Protein into a Bright Blue Probe. Chemistry and Biology, 2008, 15, 1116-1124.	6.2	269
3	Red fluorescent genetically encoded indicator for intracellular hydrogen peroxide. Nature Communications, 2014, 5, 5222.	5.8	207
4	Bright Monomeric Photoactivatable Red Fluorescent Protein for Two-Color Super-Resolution sptPALM of Live Cells. Journal of the American Chemical Society, 2010, 132, 6481-6491.	6.6	190
5	Green fluorescent proteins are light-induced electron donors. Nature Chemical Biology, 2009, 5, 459-461.	3.9	176
6	Chromophore Transformations in Red Fluorescent Proteins. Chemical Reviews, 2012, 112, 4308-4327.	23.0	173
7	Monomeric fluorescent timers that change color from blue to red report on cellular trafficking. Nature Chemical Biology, 2009, 5, 118-126.	3.9	164
8	Superresolution Imaging of Multiple Fluorescent Proteins with Highly Overlapping Emission Spectra in Living Cells. Biophysical Journal, 2011, 101, 1522-1528.	0.2	139
9	Engineering of bacterial phytochromes for near-infrared imaging, sensing, and light-control in mammals. Chemical Society Reviews, 2013, 42, 3441.	18.7	134
10	Red Fluorescent Protein with Reversibly Photoswitchable Absorbance for Photochromic FRET. Chemistry and Biology, 2010, 17, 745-755.	6.2	123
11	Far-red light photoactivatable near-infrared fluorescent proteins engineered from a bacterial phytochrome. Nature Communications, 2013, 4, 2153.	5.8	108
12	Photoactivation mechanism of PAmCherry based on crystal structures of the protein in the dark and fluorescent states. Proceedings of the National Academy of Sciences of the United States of America, 2009, 106, 21097-21102.	3.3	86
13	Flow cytometry of fluorescent proteins. Methods, 2012, 57, 318-330.	1.9	77
14	Directed molecular evolution to design advanced red fluorescent proteins. Nature Methods, 2011, 8, 1019-1026.	9.0	72
15	The First Mutant of the Aequorea victoria Green Fluorescent Protein That Forms a Red Chromophore. Biochemistry, 2008, 47, 4666-4673.	1.2	67
16	Solid state yellow and orange lasers for flow cytometry. Cytometry Part A: the Journal of the International Society for Analytical Cytology, 2008, 73A, 570-577.	1.1	58
17	Understanding Blue-to-Red Conversion in Monomeric Fluorescent Timers and Hydrolytic Degradation of Their Chromophores. Journal of the American Chemical Society, 2010, 132, 2243-2253.	6.6	51
18	New lasers for flow cytometry: filling the gaps. Nature Methods, 2007, 4, 678-679.	9.0	44

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19	A Structural Basis for Reversible Photoswitching of Absorbance Spectra in Red Fluorescent Protein rsTagRFP. Journal of Molecular Biology, 2012, 417, 144-151.	2.0	40
20	A new design for a green calcium indicator with a smaller size and a reduced number of calcium-binding sites. Scientific Reports, 2016, 6, 34447.	1.6	35
21	Novel Genetically Encoded Bright Positive Calcium Indicator NCaMP7 Based on the mNeonGreen Fluorescent Protein. International Journal of Molecular Sciences, 2020, 21, 1644.	1.8	33
22	Advances in Engineering and Application of Optogenetic Indicators for Neuroscience. Applied Sciences (Switzerland), 2019, 9, 562.	1.3	32
23	Effects of Benzo[a]pyreneâ^'Deoxyguanosine Lesions on DNA Methylation Catalyzed by EcoRII DNA Methyltransferase and on DNA Cleavage Effected by EcoRII Restriction Endonucleaseâ€. Biochemistry, 2005, 44, 1054-1066.	1.2	29
24	Near-Infrared Genetically Encoded Positive Calcium Indicator Based on GAF-FP Bacterial Phytochrome. International Journal of Molecular Sciences, 2019, 20, 3488.	1.8	28
25	NTnC-like genetically encoded calcium indicator with a positive and enhanced response and fast kinetics. Scientific Reports, 2018, 8, 15233.	1.6	24
26	Slowly Reducible Genetically Encoded Green Fluorescent Indicator for In Vivo and Ex Vivo Visualization of Hydrogen Peroxide. International Journal of Molecular Sciences, 2019, 20, 3138.	1.8	24
27	Supercontinuum white light lasers for flow cytometry. Cytometry Part A: the Journal of the International Society for Analytical Cytology, 2009, 75A, 450-459.	1.1	22
28	Green fluorescent genetically encoded calcium indicator based on calmodulin/M13-peptide from fungi. PLoS ONE, 2017, 12, e0183757.	1.1	22
29	FGCaMP7, an Improved Version of Fungi-Based Ratiometric Calcium Indicator for In Vivo Visualization of Neuronal Activity. International Journal of Molecular Sciences, 2020, 21, 3012.	1.8	17
30	Tuning the Sensitivity of Genetically Encoded Fluorescent Potassium Indicators through Structure-Guided and Genome Mining Strategies. ACS Sensors, 2022, 7, 1336-1346.	4.0	17
31	Genetically encoded calcium indicator with NTnC-like design and enhanced fluorescence contrast and kinetics. BMC Biotechnology, 2018, 18, 10.	1.7	16
32	Rapid directed molecular evolution of fluorescent proteins in mammalian cells. Protein Science, 2022, 31, 728-751.	3.1	11
33	GAF-CaMP3–sfGFP, An Enhanced Version of the Near-Infrared Genetically Encoded Positive Phytochrome-Based Calcium Indicator for the Visualization of Neuronal Activity. International Journal of Molecular Sciences, 2020, 21, 6883.	1.8	10
34	LSSmScarlet, dCyRFP2s, dCyOFP2s and CRISPRed2s, Genetically Encoded Red Fluorescent Proteins with a Large Stokes Shift. International Journal of Molecular Sciences, 2021, 22, 12887.	1.8	9
35	FRCaMP, a Red Fluorescent Genetically Encoded Calcium Indicator Based on Calmodulin from Schizosaccharomyces Pombe Fungus. International Journal of Molecular Sciences, 2021, 22, 111.	1.8	7
36	Determination of two-photon photoactivation rates of fluorescent proteins. Physical Chemistry Chemical Physics, 2013, 15, 14868.	1.3	6

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37	DNA Duplexes Containing Altered Sugar Residues as Probes ofEcoRII andMval Endonuclease Interactions with Sugar-Phosphate Backbone. Journal of Biomolecular Structure and Dynamics, 2000, 17, 857-870.	2.0	5
38	The mRubyFT Protein, Genetically Encoded Blue-to-Red Fluorescent Timer. International Journal of Molecular Sciences, 2022, 23, 3208.	1.8	5
39	The whither of bacteriophytochromeâ€based nearâ€infrared fluorescent proteins: Insights from twoâ€photon absorption spectroscopy. Journal of Biophotonics, 2019, 12, e201800353.	1.1	4
40	The rotational order–disorder structure of the reversibly photoswitchable red fluorescent protein rsTagRFP. Acta Crystallographica Section D: Biological Crystallography, 2014, 70, 31-39.	2.5	4
41	Investigation of restriction endonuclease EcoRII complex with DNA in solution by FTIR spectroscopy. Russian Journal of General Chemistry, 2008, 78, 1103-1109.	0.3	3
42	Visualizing the Receptor Assembly Into Clathrin-coated Pits with Super-resolution Two-color Palm and sptPALM. Biophysical Journal, 2009, 96, 385a.	0.2	1
43	Resolution of the EcoRII restriction endonuclease-DNA complex structure in solution using fluorescence spectroscopy. Biophysical Chemistry, 2008, 138, 107-114.	1.5	Ο