Nicholas Rhind

List of Publications by Year
in descending order

60 papers	21,831 citations	126708 33 h-index	138251 58 g-index
69 all docs	69 docs citations	69 times ranked	31382 citing authors

#	Article	IF	Citations
1	Full-length transcriptome assembly from RNA-Seq data without a reference genome. Nature Biotechnology, 2011, 29, 644-652.	9.4	17,264
2	Cdc25 Mitotic Inducer Targeted by Chk1 DNA Damage Checkpoint Kinase. Science, 1997, 277, 1495-1497.	6.0	515
3	Comparative Functional Genomics of the Fission Yeasts. Science, 2011, 332, 930-936.	6.0	458
4	Basic methods for fission yeast. Yeast, 2006, 23, 173-183.	0.8	457
5	DNA Replication Timing. Cold Spring Harbor Perspectives in Biology, 2013, 5, a010132-a010132.	2.3	278
6	Chromosome Mis-segregation Generates Cell-Cycle-Arrested Cells with Complex Karyotypes that Are Eliminated by the Immune System. Developmental Cell, 2017, 41, 638-651.e5.	3.1	263
7	DNA Replication Origins Fire Stochastically in Fission Yeast. Molecular Biology of the Cell, 2006, 17, 308-316.	0.9	176
8	Mitotic DNA damage and replication checkpoints in yeast. Current Opinion in Cell Biology, 1998, 10, 749-758.	2.6	159
9	Signaling Pathways that Regulate Cell Division. Cold Spring Harbor Perspectives in Biology, 2012, 4, a005942-a005942.	2.3	129
10	A single Argonaute protein mediates both transcriptional and posttranscriptional silencing in Schizosaccharomyces pombe. Genes and Development, 2004, 18, 2359-2367.	2.7	128
11	DNA replication timing: random thoughts about origin firing. Nature Cell Biology, 2006, 8, 1313-1316.	4.6	116
12	Modeling genomeâ€wide replication kinetics reveals a mechanism for regulation of replication timing. Molecular Systems Biology, 2010, 6, 404.	3.2	113
13	Tyrosine Phosphorylation of Cdc2 Is Required for the Replication Checkpoint in <i>Schizosaccharomyces pombe</i>. Molecular and Cellular Biology, 1998, 18, 3782-3787.	1.1	109
14	xol-1 acts as an early switch in the C. elegans male/hermaphrodite decision. Cell, 1995, 80, 71-82.	13.5	94
15	Replication timing and its emergence from stochastic processes. Trends in Genetics, 2012, 28, 374-381.	2.9	87
16	The Intra-S Checkpoint Responses to DNA Damage. Genes, 2017, 8, 74.	1.0	87
17	Genome-wide identification and characterization of replication origins by deep sequencing. Genome Biology, 2012, 13, R27.	13.9	85
18	Size-Dependent Expression of the Mitotic Activator Cdc25 Suggests a Mechanism of Size Control in Fission Yeast. Current Biology, 2017, 27, 1491-1497.e4.	1.8	84

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19	The Hsk1(Cdc7) Replication Kinase Regulates Origin Efficiency. Molecular Biology of the Cell, 2008, 19, 5550-5558.	0.9	81
20	Replication timing is regulated by the number of MCMs loaded at origins. Genome Research, 2015, 25, 1886-1892.	2.4	80
21	Evolutionary divergence of intrinsic and <i>trans</i>-regulated nucleosome positioning sequences reveals plastic rules for chromatin organization. Genome Research, 2011, 21, 1851-1862.	2.4	74
22	Roles of the Mitotic Inhibitors Wee1 and Mik1 in the G 2 DNA Damage and Replication Checkpoints. Molecular and Cellular Biology, 2001, 21, 1499-1508.	1.1	73
23	The Fission Yeast Rad32 (Mre11)-Rad50-Nbs1 Complex Is Required for the S-Phase DNA Damage Checkpoint. Molecular and Cellular Biology, 2003, 23, 6564-6573.	1.1	70
24	Reconciling stochastic origin firing with defined replication timing. Chromosome Research, 2010, 18, 35-43.	1.0	69
25	Regulation of DNA replication by the S-phase DNA damage checkpoint. Cell Division, 2009, 4, 13.	1.1	66
26	The Schizosaccharomyces pombe S-Phase Checkpoint Differentiates Between Different Types of DNA Damage. Genetics, 1998, 149, 1729-1737.	1.2	63
27	Checkpoints: It takes more than time to heal some wounds. Current Biology, 2000, 10, R908-R911.	1.8	59
28	In vivo labeling of fission yeast DNA with thymidine and thymidine analogs. Methods, 2004, 33, 213-219.	1.9	59
29	Genome-wide mapping of human DNA replication by optical replication mapping supports a stochastic model of eukaryotic replication. Molecular Cell, 2021, 81, 2975-2988.e6.	4.5	57
30	The DNA Replication Checkpoint Directly Regulates MBF-Dependent G₁/S Transcription. Molecular and Cellular Biology, 2008, 28, 5977-5985.	1.1	47
31	Global increase in replication fork speed during a p57 ^{KIP2} -regulated erythroid cell fate switch. Science Advances, 2017, 3, e1700298.	4.7	44
32	Replication fork slowing and stalling are distinct, checkpoint-independent consequences of replicating damaged DNA. PLoS Genetics, 2017, 13, e1006958.	1.5	43
33	Mre11 Nuclease Activity and Ctp1 Regulate Chk1 Activation by Rad3^{ATR} and Tel1^{ATM} Checkpoint Kinases at Double-Strand Breaks. Molecular and Cellular Biology, 2011, 31, 573-583.	1.1	38
34	Mus81, Rhp51(Rad51), and Rqh1 Form an Epistatic Pathway Required for the S-Phase DNA Damage Checkpoint. Molecular Biology of the Cell, 2009, 20, 819-833.	0.9	37
35	How and why multiple MCMs are loaded at origins of DNA replication. BioEssays, 2016, 38, 613-617.	1.2	26
36	Transcriptome-wide Interrogation of the Functional Intronome by Spliceosome Profiling. Cell, 2018, 173, 1031-1044.e13.	13.5	26

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37	The capacity of origins to load MCM establishes replication timing patterns. PLoS Genetics, 2021, 17, e1009467.	1.5	22
38	An estradiolâ€inducible promoter enables fast, graduated control of gene expression in fission yeast. Yeast, 2017, 34, 323-334.	0.8	20
39	Cell-size control. Current Biology, 2021, 31, R1414-R1420.	1.8	16
40	Checkpoint regulation of replication forks: global or local?. Biochemical Society Transactions, 2013, 41, 1701-1705.	1.6	15
41	Cdc2 Tyrosine Phosphorylation is Not Required for the S-Phase DNA Damage Checkpoint in Fission Yeast. Cell Cycle, 2006, 5, 2495-2500.	1.3	14
42	Identification of S-phase DNA damage-response targets in fission yeast reveals conservation of damage-response networks. Proceedings of the National Academy of Sciences of the United States of America, 2016, 113, E3676-E3685.	3.3	13
43	The Role of MRN in the S-Phase DNA Damage Checkpoint Is Independent of Its Ctp1-dependent Roles in Double-Strand Break Repair and Checkpoint Signaling. Molecular Biology of the Cell, 2009, 20, 2096-2107.	0.9	12
44	Changing of the Guard: How ATM Hands Off DNA Double-Strand Break Signaling to ATR. Molecular Cell, 2009, 33, 672-674.	4.5	10
45	Fission yeast cells grow approximately exponentially. Cell Cycle, 2019, 18, 869-879.	1.3	10
46	Studying G2 DNA Damage Checkpoints Using the Fission Yeast Schizosaccharomyces pombe. Methods in Molecular Biology, 2011, 782, 1-12.	0.4	10
47	The Role of Specific Checkpoint-Induced S-Phase Transcripts in Resistance to Replicative Stress. PLoS ONE, 2009, 4, e6944.	1.1	9
48	Cell Size Control via an Unstable Accumulating Activator and the Phenomenon of Excess Mitotic Delay. BioEssays, 2018, 40, 1700184.	1.2	7
49	Incorporation of Thymidine Analogs for Studying Replication Kinetics in Fission Yeast. Methods in Molecular Biology, 2009, 521, 509-515.	0.4	6
50	An intrinsic checkpoint model for regulation of replication origins. Cell Cycle, 2008, 7, 2619-2620.	1.3	5
51	The Fission Yeast Rad32(Mre11)â€“Rad50â€“Nbs1 Complex Acts Both Upstream and Downstream of Checkpoint Signaling in the S-Phase DNA Damage Checkpoint. Genetics, 2010, 184, 887-897.	1.2	5
52	Discovery of genes involved in mitosis, cell division, cell wall integrity and chromosome segregation through construction of <i>Schizosaccharomyces pombe</i> deletion strains. Yeast, 2016, 33, 507-517.	0.8	5
53	Studying S-Phase DNA Damage Checkpoints Using the Fission Yeast Schizosaccharomyces pombe. Methods in Molecular Biology, 2011, 782, 13-21.	0.4	4
54	The three most important things about origins: location, location, location. Molecular Systems Biology, 2014, 10, 723.	3.2	3

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55	DNA Damage Checkpoint Control of Mitosis in Fission Yeast. Cold Spring Harbor Symposia on Quantitative Biology, 2000, 65, 353-360.	2.0	3
56	The fission yeast S-phase cyclin Cig2 can drive mitosis. Genetics, 2021, 217, 1-12.	1.2	2
57	Incorporation of Thymidine Analogs for Studying Replication Kinetics in Fission Yeast. Methods in Molecular Biology, 2015, 1300, 99-104.	0.4	1
58	f = m*a: A Framework for Investigating the Regulation of Replication Timing. Genes, 2022, 13, 249.	1.0	1
59	Global Increase in Replication Fork Speed during a p57KIP2-Regulated Erythroid Cell Fate Switch. Blood, 2016, 128, 698-698.	0.6	0
60	Mapping replication forks, one replicon at a time. Molecular Cell, 2022, 82, 1246-1248.	4.5	0