Elizabeth J Harry

List of Publications by Year
in descending order

90 papers	4,955 citations	66343 42 h-index	98798 67 g-index
92 all docs	92 docs citations	92 times ranked	4946 citing authors

#	Article	IF	Citations
1	Inhibition of Dermatophyte Fungi by Australian Jarrah Honey. Pathogens, 2021, 10, 194.	2.8	6
2	Factors affecting the production and measurement of hydrogen peroxide in honey samples. Access Microbiology, 2021, 3, 000198.	0.5	4
3	A Newly Identified Prophage Gene, <i>ymfM</i> , Causes SOS-Inducible Filamentation in Escherichia coli. Journal of Bacteriology, 2021, 203, .	2.2	7
4	Cataloguing the small RNA content of honey using next generation sequencing. Food Chemistry Molecular Sciences, 2021, 2, 100014.	2.1	7
5	Synthesis and biological evaluation of 3,5-substituted pyrazoles as possible antibacterial agents. Bioorganic and Medicinal Chemistry, 2021, 48, 116401.	3.0	4
6	Heritable nanosilver resistance in priority pathogen: a unique genetic adaptation and comparison with ionic silver and antibiotics. Nanoscale, 2020, 12, 2384-2392.	5.6	29
7	Characterizing the Mechanism of Action of an Ancient Antimicrobial, Manuka Honey, against Pseudomonas aeruginosa Using Modern Transcriptomics. MSystems, 2020, 5, .	3.8	30
8	Uncovering novel susceptibility targets to enhance the efficacy of third-generation cephalosporins against ESBL-producing uropathogenic Escherichia coli. Journal of Antimicrobial Chemotherapy, 2020, 75, 1415-1423.	3.0	7
9	The novel E. coli cell division protein, YtfB, plays a role in eukaryotic cell adhesion. Scientific Reports, 2020, 10, 6745.	3.3	3
10	FtsZ as an Antibacterial Target: Status and Guidelines for Progressing This Avenue. ACS Infectious Diseases, 2019, 5, 1279-1294.	3.8	47
11	The ParB homologs, Spo0J and Noc, together prevent premature midcell Z ring assembly when the early stages of replication are blocked in Bacillus subtilis. Molecular Microbiology, 2019, 112, 766-784.	2.5	11
12	Honey can inhibit and eliminate biofilms produced by Pseudomonas aeruginosa. Scientific Reports, 2019, 9, 18160.	3.3	63
13	Analysis of FtsZ Crystal Structures Towards a New Target for Antibiotics. Australian Journal of Chemistry, 2019, 72, 184-193.	0.9	9
14	A cost-effective colourimetric assay for quantifying hydrogen peroxide in honey. Access Microbiology, 2019, 1, e000065.	0.5	11
15	Nanosilver and the microbiological activity of the particulate solids versus the leached soluble silver. Nanotoxicology, 2018, 12, 263-273.	3.0	23
16	High-throughput sequencing of sorted expression libraries reveals inhibitors of bacterial cell division. BMC Genomics, 2018, 19, 781.	2.8	6
17	DNA condensation in live <i>E. coli</i> provides evidence for transertion. Molecular BioSystems, 2017, 13, 677-680.	2.9	7
18	Immobilization Techniques of Bacteria for Live Super-resolution Imaging Using Structured Illumination Microscopy. Methods in Molecular Biology, 2017, 1535, 197-209.	0.9	8

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19	Widespread and Indiscriminate Nanosilver Use: Genuine Potential for Microbial Resistance. ACS Nano, 2017, 11, 3438-3445.	14.6	77
20	We Are What We Eat: True for Bacteria Too. Frontiers for Young Minds, 2017, 5, .	0.8	0
21	Metabolic Adaptations of Uropathogenic E. coli in the Urinary Tract. Frontiers in Cellular and Infection Microbiology, 2017, 7, 241.	3.9	93
22	Coordination of Chromosome Segregation and Cell Division in Staphylococcus aureus. Frontiers in Microbiology, 2017, 8, 1575.	3.5	29
23	Rifampicin-Manuka Honey Combinations Are Superior to Other Antibiotic-Manuka Honey Combinations in Eradicating Staphylococcus aureus Biofilms. Frontiers in Microbiology, 2017, 8, 2653.	3.5	37
24	A longitudinal study of the diabetic skin and wound microbiome. PeerJ, 2017, 5, e3543.	2.0	93
25	Therapeutic Manuka Honey: No Longer So Alternative. Frontiers in Microbiology, 2016, 7, 569.	3.5	128
26	The Antibacterial Activity of Australian Leptospermum Honey Correlates with Methylglyoxal Levels. PLoS ONE, 2016, 11, e0167780.	2.5	61
27	Understanding, Monitoring, and Controlling Biofilm Growth in Drinking Water Distribution Systems. Environmental Science & Technology, 2016, 50, 8954-8976.	10.0	302
28	You Are What You Eat: Metabolic Control of Bacterial Division. Trends in Microbiology, 2016, 24, 181-189.	7.7	26
29	Connecting the dots of the bacterial cell cycle: Coordinating chromosome replication and segregation with cell division. Seminars in Cell and Developmental Biology, 2016, 53, 2-9.	5.0	42
30	Cryptococcus Strains with Different Pathogenic Potentials Have Diverse Protein Secretomes. Eukaryotic Cell, 2015, 14, 554-563.	3.4	28
31	CetZ tubulin-like proteins control archaeal cell shape. Nature, 2015, 519, 362-365.	27.8	138
32	You are what you secrete: extracellular proteins and virulence in Cryptococcus. Microbiology Australia, 2015, 36, 93.	0.4	2
33	Division site positioning in bacteria: one size does not fit all. Frontiers in Microbiology, 2014, 5, 19.	3.5	78
34	Manuka-type honeys can eradicate biofilms produced by<i>Staphylococcus aureus</i>strains with different biofilm-forming abilities. PeerJ, 2014, 2, e326.	2.0	122
35	Super-resolution Imaging of the Cytokinetic Z Ring in Live Bacteria Using Fast 3D-Structured Illumination Microscopy (f3D-SIM). Journal of Visualized Experiments, 2014, , 51469.	0.3	14
36	Structure and function of a spectrin-like regulator of bacterial cytokinesis. Nature Communications, 2014, 5, 5421.	12.8	41

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37	A beacon for bacterial tubulin. Nature, 2014, 516, 175-176.	27.8	0
38	Coordinating Bacterial Cell Division with Nutrient Availability: a Role for Glycolysis. MBio, 2014, 5, e00935-14.	4.1	93
39	Dinuclear ruthenium(<scp>ii</scp>) antimicrobial agents that selectively target polysomes in vivo. Chemical Science, 2014, 5, 685-693.	7.4	48
40	Discovery of Lead Compounds Targeting the Bacterial Sliding Clamp Using a Fragment-Based Approach. Journal of Medicinal Chemistry, 2014, 57, 2799-2806.	6.4	49
41	DNA Replication Is the Target for the Antibacterial Effects of Nonsteroidal Anti-Inflammatory Drugs. Chemistry and Biology, 2014, 21, 481-487.	6.0	102
42	Antibiotic-specific differences in the response of Staphylococcus aureus to treatment with antimicrobials combined with manuka honey. Frontiers in Microbiology, 2014, 5, 779.	3.5	44
43	Analysis of the flavonoid component of bioactive New Zealand mÄnuka (Leptospermum scoparium) honey and the isolation, characterisation and synthesis of an unusual pyrrole. Food Chemistry, 2013, 141, 1772-1781.	8.2	68
44	Identifying how bacterial cells find their middle: a new perspective. Molecular Microbiology, 2013, 87, 231-234.	2.5	17
45	Synergism between Medihoney and Rifampicin against Methicillin-Resistant Staphylococcus aureus (MRSA). PLoS ONE, 2013, 8, e57679.	2.5	91
46	The Effect of New Zealand Kanuka, Manuka and Clover Honeys on Bacterial Growth Dynamics and Cellular Morphology Varies According to the Species. PLoS ONE, 2013, 8, e55898.	2.5	88
47	Harnessing Single Cell Sorting to Identify Cell Division Genes and Regulators in Bacteria. PLoS ONE, 2013, 8, e60964.	2.5	27
48	The Min System and Nucleoid Occlusion Are Not Required for Identifying the Division Site in Bacillus subtilis but Ensure Its Efficient Utilization. PLoS Genetics, 2012, 8, e1002561.	3.5	79
49	3D-SIM Super Resolution Microscopy Reveals a Bead-Like Arrangement for FtsZ and the Division Machinery: Implications for Triggering Cytokinesis. PLoS Biology, 2012, 10, e1001389.	5.6	186
50	Specific non-peroxide antibacterial effect of manuka honey on the Staphylococcus aureus proteome. International Journal of Antimicrobial Agents, 2012, 40, 43-50.	2.5	58
51	Time-Course Proteome Analysis Reveals the Dynamic Response of Cryptococcus gattii Cells to Fluconazole. PLoS ONE, 2012, 7, e42835.	2.5	17
52	Super-resolution imaging of the bacterial cytokinetic protein FtsZ. Micron, 2011, 42, 336-341.	2.2	54
53	A simple plasmid-based system that allows rapid generation of tightly controlled gene expression in Staphylococcus aureus. Microbiology (United Kingdom), 2011, 157, 666-676.	1.8	40
54	Influence of the nucleoid and the early stages of DNA replication on positioning the division site in Bacillus subtilis. Molecular Microbiology, 2010, 76, 634-647.	2.5	46

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55	Essential Biological Processes of an Emerging Pathogen: DNA Replication, Transcription, and Cell Division in <i>Acinetobacter</i> spp. Microbiology and Molecular Biology Reviews, 2010, 74, 273-297.	6.6	68
56	Effects of oriC relocation on control of replication initiation in Bacillus subtilis. Microbiology (United Kingdom), 2009, 155, 3070-3082.	1.8	4
57	Lateral FtsZ association and the assembly of the cytokinetic Z ring in bacteria. Molecular Microbiology, 2009, 74, 1004-1017.	2.5	68
58	Cell-division inhibitors: new insights for future antibiotics. Nature Reviews Drug Discovery, 2008, 7, 324-338.	46.4	221
59	The divisomal protein DivIB contains multiple epitopes that mediate its recruitment to incipient division sites. Molecular Microbiology, 2008, 67, 1143-1155.	2.5	12
60	A new assembly pathway for the cytokinetic Z ring from a dynamic helical structure in vegetatively growing cells of Bacillus subtilis. Molecular Microbiology, 2007, 64, 487-499.	2.5	85
61	Bacterial Cell Division: The Mechanism and Its Precison. International Review of Cytology, 2006, 253, 27-94.	6.2	158
62	Improved 2-DE of microorganisms after acidic extraction. Electrophoresis, 2006, 27, 1630-1640.	2.4	29
63	Requirement for the Cell Division Protein DivIB in Polar Cell Division and Engulfment during Sporulation in Bacillus subtilis. Journal of Bacteriology, 2006, 188, 7677-7685.	2.2	12
64	Trapping of a Spiral-Like Intermediate of the Bacterial Cytokinetic Protein FtsZ. Journal of Bacteriology, 2006, 188, 1680-1690.	2.2	46
65	A streamlined approach to high-throughput proteomics. Expert Review of Proteomics, 2005, 2, 173-185.	3.0	4
66	Bacillus subtilisYabA is involved in determining the timing and synchrony of replication initiation. FEMS Microbiology Letters, 2005, 247, 73-79.	1.8	39
67	Cell Division in<i>Bacillus subtilis</i>: FtsZ and FtsA Association Is Z-Ring Independent, and FtsA Is Required for Efficient Midcell Z-Ring Assembly. Journal of Bacteriology, 2005, 187, 6536-6544.	2.2	79
68	The midcell replication factory in Bacillus subtilis is highly mobile: implications for coordinating chromosome replication with other cell cycle events. Molecular Microbiology, 2004, 54, 452-463.	2.5	56
69	Cell division protein DivIB influences the Spo0J/Soj system of chromosome segregation in Bacillus subtilis. Molecular Microbiology, 2004, 55, 349-367.	2.5	25
70	Early targeting of Min proteins to the cell poles in germinated spores of Bacillus subtilis : evidence for division apparatusâ€independent recruitment of Min proteins to the division site. Molecular Microbiology, 2003, 47, 37-48.	2.5	58
71	Increasing the Ratio of Soj to Spo0J Promotes Replication Initiation in Bacillus subtilis. Journal of Bacteriology, 2003, 185, 6316-6324.	2.2	47
72	The Min system is not required for precise placement of the midcell Z ring in Bacillus subtilis. EMBO Reports, 2002, 3, 1163-1167.	4.5	51

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73	The Bacillus subtilis cell division proteins FtsL and DivIC are intrinsically unstable and do not interact with one another in the absence of other septasomal components. Molecular Microbiology, 2002, 44, 663-674.	2.5	42
74	Coordinating DNA replication with cell division:Lessons from outgrowing spores. Biochimie, 2001, 83, 75-81.	2.6	18
75	Bacterial cell division: regulating Z-ring formation. Molecular Microbiology, 2001, 40, 795-803.	2.5	96
76	Role of penicillin-binding protein PBP 2B in assembly and functioning of the division machinery of Bacillus subtilis. Molecular Microbiology, 2000, 35, 299-311.	2.5	113
77	Mid-cell Z ring assembly in the absence of entry into the elongation phase of the round of replication in bacteria: co-ordinating chromosome replication with cell division. Molecular Microbiology, 2000, 38, 423-434.	2.5	47
78	Septal Localization of the Membrane-Bound Division Proteins ofBacillus subtilis DivIB and DivIC Is Codependent Only at High Temperatures and Requires FtsZ. Journal of Bacteriology, 2000, 182, 3607-3611.	2.2	37
79	Co-ordinating DNA replication with cell division in bacteria: a link between the early stages of a round of replication and mid-cell Z ring assembly. Molecular Microbiology, 1999, 33, 33-40.	2.5	70
80	Characterization of the essential cell division geneftsLâ€ƒ(yllDâ€Š) ofBacillus subtilisand its role in the assembly of the division apparatus. Molecular Microbiology, 1998, 29, 593-604.	2.5	112
81	Illuminating the force: Bacterial mitosis?. Trends in Microbiology, 1997, 5, 295-297.	7.7	6
82	The membrane-bound cell division protein DivIB is localized to the division site inBacillus subtilis. Molecular Microbiology, 1997, 25, 275-283.	2.5	56
83	The Bacillus subtilis division protein DivIC is a highly abundant membraneâ€bound protein that localizes to the division site. Molecular Microbiology, 1997, 26, 1047-1055.	2.5	45
84	Use of immunofluorescence to visualize cell-specific gene expression during sporulation in Bacillus subtilis. Journal of Bacteriology, 1995, 177, 3386-3393.	2.2	181
85	Visualization of the subcellular location of sporulation proteins in Bacillus subtilis using immunofluorescence microscopy. Molecular Microbiology, 1995, 18, 459-470.	2.5	149
86	Extracellular signal protein triggering the proteolytic activation of a developmental transcription factor in B. subtilis. Cell, 1995, 83, 219-226.	28.9	109
87	Expression of divIB of Bacillus subtilis during vegetative growth. Journal of Bacteriology, 1994, 176, 1172-1179.	2.2	10
88	Conservation of the 168 divIB gene in Bacillus subtilis W23 and B. licheniformis, and evidence for homology to ftsQ of Escherichia coli. Gene, 1994, 147, 85-89.	2.2	19
89	Characterization of mutations in divlB of Bacillus subtilis and cellular localization of the DivlB protein. Molecular Microbiology, 1993, 7, 611-621.	2.5	42
90	Cloning and expression of a Bacillus subtilis division initiation gene for which a homolog has not been identified in another organism. Journal of Bacteriology, 1989, 171, 6835-6839.	2.2	40