## EncarnaciÃ<sup>3</sup>n MartÃ-nez-Salas

List of Publications by Year in descending order

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Version: 2024-02-01



#	Article	IF	CITATIONS
1	Autosomal Recessive Cerebellar Atrophy and Spastic Ataxia in Patients With Pathogenic Biallelic Variants in GEMIN5. Frontiers in Cell and Developmental Biology, 2022, 10, 783762.	3.7	10
2	Picornavirus translation strategies. FEBS Open Bio, 2022, 12, 1125-1141.	2.3	21
3	Functional and structural deficiencies of Gemin5 variants associated with neurological disorders. Life Science Alliance, 2022, 5, e202201403.	2.8	7
4	Identification of RNA-Binding Proteins Associated to RNA Structural Elements. Methods in Molecular Biology, 2021, 2323, 109-119.	0.9	1
5	Uncovering targets of the Leader protease: Linking RNA â€mediated pathways and antiviral defense. Wiley Interdisciplinary Reviews RNA, 2021, 12, e1645.	6.4	9
6	RNA-Binding Proteins at the Host-Pathogen Interface Targeting Viral Regulatory Elements. Viruses, 2021, 13, 952.	3.3	15
7	The RBS1 domain of Gemin5 is intrinsically unstructured and interacts with RNA through conserved Arg and aromatic residues. RNA Biology, 2021, 18, 496-506.	3.1	7
8	Structural insights of the pre-let-7 interaction with LIN28B. Nucleosides, Nucleotides and Nucleic Acids, 2021, 40, 1-19.	1.1	2
9	Structural basis for the dimerization of Gemin5 and its role in protein recruitment and translation control. Nucleic Acids Research, 2020, 48, 788-801.	14.5	19
10	MDA5 cleavage by the Leader protease of foot-and-mouth disease virus reveals its pleiotropic effect against the host antiviral response. Cell Death and Disease, 2020, 11, 718.	6.3	15
11	RNA-protein coevolution study of Gemin5 uncovers the role of the PXSS motif of RBS1 domain for RNA binding. RNA Biology, 2020, 17, 1331-1341.	3.1	10
12	Emerging Roles of Gemin5: From snRNPs Assembly to Translation Control. International Journal of Molecular Sciences, 2020, 21, 3868.	4.1	24
13	Thermostability of the Foot-and-Mouth Disease Virus Capsid Is Modulated by Lethal and Viability-Restoring Compensatory Amino Acid Substitutions. Journal of Virology, 2019, 93, .	3.4	9
14	Impact of RNA–Protein Interaction Modes on Translation Control: The Versatile Multidomain Protein Gemin5. BioEssays, 2019, 41, e1800241.	2.5	20
15	A Combined ELONA-(RT)qPCR Approach for Characterizing DNA and RNA Aptamers Selected against PCBP-2. Molecules, 2019, 24, 1213.	3.8	14
16	Genome Organisation, Translation and Replication of Foot-and-Mouth Disease Virus RNA. , 2019, , 19-52.		7
17	Rab1b and ARF5 are novel RNA-binding proteins involved in FMDV IRES–driven RNA localization. Life Science Alliance, 2019, 2, e201800131.	2.8	14
18	Ribosome-dependent conformational flexibility changes and RNA dynamics of IRES domains revealed by differential SHAPE. Scientific Reports, 2018, 8, 5545.	3.3	18

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19	Deconstructing internal ribosome entry site elements: an update of structural motifs and functional divergences. Open Biology, 2018, 8, 180155.	3.6	15
20	The landscape of the non-canonical RNA-binding site of Gemin5 unveils a feedback loop counteracting the negative effect on translation. Nucleic Acids Research, 2018, 46, 7339-7353.	14.5	23
21	Innate immune sensor LGP2 is cleaved by the Leader protease of foot-and-mouth disease virus. PLoS Pathogens, 2018, 14, e1007135.	4.7	35
22	In-cell SHAPE uncovers dynamic interactions between the untranslated regions of the foot-and-mouth disease virus RNA. Nucleic Acids Research, 2017, 45, gkw795.	14.5	28
23	G3 <scp>BP</scp> 1 interacts directly with the <scp>FMDV IRES</scp> and negatively regulates translation. FEBS Journal, 2017, 284, 3202-3217.	4.7	42
24	Insights into Structural and Mechanistic Features of Viral IRES Elements. Frontiers in Microbiology, 2017, 8, 2629.	3.5	100
25	Genome Organisation, Translation and Replication of Foot-and-mouth Disease Virus RNA. , 2017, , 13-42.		6
26	IRES Elements: Issues, Controversies and Evolutionary Perspectives. , 2016, , 547-564.		2
27	RNAiFold2T: Constraint Programming design of thermo-IRES switches. Bioinformatics, 2016, 32, i360-i368.	4.1	8
28	The RNA-binding protein Gemin5 binds directly to the ribosome and regulates global translation. Nucleic Acids Research, 2016, 44, 8335-8351.	14.5	54
29	Designing synthetic RNAs to determine the relevance of structural motifs in picornavirus IRES elements. Scientific Reports, 2016, 6, 24243.	3.3	8
30	Fingerprinting the junctions of RNA structure by an open-paddlewheel diruthenium compound. Rna, 2016, 22, 330-338.	3.5	19
31	Modeling Three-Dimensional Structural Motifs of Viral IRES. Journal of Molecular Biology, 2016, 428, 767-776.	4.2	23
32	Gemin5: A Multitasking RNA-Binding Protein Involved in Translation Control. Biomolecules, 2015, 5, 528-544.	4.0	38
33	Structural insights into viral IRES-dependent translation mechanisms. Current Opinion in Virology, 2015, 12, 113-120.	5.4	132
34	Picornavirus IRES elements: RNA structure and host protein interactions. Virus Research, 2015, 206, 62-73.	2.2	110
35	Local RNA flexibility perturbation of the IRES element induced by a novel ligand inhibits viral RNA translation. RNA Biology, 2015, 12, 555-568.	3.1	25
36	RNA–protein interaction methods to study viral IRES elements. Methods, 2015, 91, 3-12.	3.8	24

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37	Functional and Structural Analysis of Maize Hsp101 IRES. PLoS ONE, 2014, 9, e107459.	2.5	11
38	Identification of novel non-canonical RNA-binding sites in Gemin5 involved in internal initiation of translation. Nucleic Acids Research, 2014, 42, 5742-5754.	14.5	47
39	Magnesiumâ€dependent folding of a picornavirus <scp>IRES</scp> element modulates <scp>RNA</scp> conformation and elF4G interaction. FEBS Journal, 2014, 281, 3685-3700.	4.7	26
40	Increased Replicative Fitness Can Lead to Decreased Drug Sensitivity of Hepatitis C Virus. Journal of Virology, 2014, 88, 12098-12111.	3.4	74
41	Enhanced IRES activity by the 3′UTR element determines the virulence of FMDV isolates. Virology, 2014, 448, 303-313.	2.4	28
42	RNA-Binding Proteins Impacting on Internal Initiation of Translation. International Journal of Molecular Sciences, 2013, 14, 21705-21726.	4.1	50
43	Evolutionary conserved motifs constrain the RNA structure organization of picornavirus IRES. FEBS Letters, 2013, 587, 1353-1358.	2.8	16
44	Gemin5 promotes IRES interaction and translation control through its C-terminal region. Nucleic Acids Research, 2013, 41, 1017-1028.	14.5	55
45	Using RNA inverse folding to identify IRES-like structural subdomains. RNA Biology, 2013, 10, 1842-1852.	3.1	20
46	Gemin5 proteolysis reveals a novel motif to identify L protease targets. Nucleic Acids Research, 2012, 40, 4942-4953.	14.5	47
47	Alternative Mechanisms to Initiate Translation in Eukaryotic mRNAs. Comparative and Functional Genomics, 2012, 2012, 1-12.	2.0	45
48	RNA Structural Elements of Hepatitis C Virus Controlling Viral RNA Translation and the Implications for Viral Pathogenesis. Viruses, 2012, 4, 2233-2250.	3.3	29
49	Exploring IRES Region Accessibility by Interference of Foot-and-Mouth Disease Virus Infectivity. PLoS ONE, 2012, 7, e41382.	2.5	12
50	Riboproteomic Approaches to Understanding IRES Elements. , 2012, , 103-118.		0
51	Structural analysis provides insights into the modular organization of picornavirus IRES. Virology, 2011, 409, 251-261.	2.4	46
52	Structural basis for the biological relevance of the invariant apical stem in IRES-mediated translation. Nucleic Acids Research, 2011, 39, 8572-8585.	14.5	58
53	Tailoring the switch from IRES-dependent to 5'-end-dependent translation with the RNase P ribozyme. Rna, 2010, 16, 852-862.	3.5	4
54	Insights into the Biology of IRES Elements through Riboproteomic Approaches. Journal of Biomedicine and Biotechnology, 2010, 2010, 1-12.	3.0	57

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55	A novel role for Gemin5 in mRNA translation. Nucleic Acids Research, 2009, 37, 582-590.	14.5	92
56	Rescue of internal initiation of translation by RNA complementation provides evidence for a distribution of functions between individual IRES domains. Virology, 2009, 388, 221-229.	2.4	21
57	Relevance of RNA structure for the activity of picornavirus IRES elements. Virus Research, 2009, 139, 172-182.	2.2	104
58	Preface. Virus Research, 2009, 139, 135-136.	2.2	0
59	Riboproteomic analysis of polypeptides interacting with the internal ribosomeâ€entry site element of footâ€endâ€mouth disease viral RNA. Proteomics, 2008, 8, 4782-4790.	2.2	60
60	Susceptibility to viral infection is enhanced by stable expression of 3A or 3AB proteins from foot-and-mouth disease virus. Virology, 2008, 380, 34-45.	2.4	17
61	Internal translation initiation on the footâ€∎ndâ€mouth disease virus IRES is affected by ribosomal stalk conformation. FEBS Letters, 2008, 582, 3029-3032.	2.8	18
62	The impact of RNA structure on picornavirus IRES activity. Trends in Microbiology, 2008, 16, 230-237.	7.7	91
63	New insights into internal ribosome entry site elements relevant for viral gene expression. Journal of General Virology, 2008, 89, 611-626.	2.9	120
64	In vivo footprint of a picornavirus internal ribosome entry site reveals differences in accessibility to specific RNA structural elements. Journal of General Virology, 2007, 88, 3053-3062.	2.9	19
65	Differential factor requirement to assemble translation initiation complexes at the alternative start codons of foot-and-mouth disease virus RNA. Rna, 2007, 13, 1366-1374.	3.5	79
66	Characterization of a cyanobacterial RNase P ribozyme recognition motif in the IRES of foot-and-mouth disease virus reveals a unique structural element. Rna, 2007, 13, 849-859.	3.5	34
67	Foot-and-mouth disease virus infection induces proteolytic cleavage of PTB, eIF3a,b, and PABP RNA-binding proteins. Virology, 2007, 364, 466-474.	2.4	62
68	Evidence of reciprocal tertiary interactions between conserved motifs involved in organizing RNA structure essential for internal initiation of translation. Rna, 2006, 12, 223-234.	3.5	78
69	The 3′ end of the foot-and-mouth disease virus genome establishes two distinct long-range RNA–RNA interactions with the 5′ end region. Journal of General Virology, 2006, 87, 3013-3022.	2.9	104
70	Cap-independent translation of maize Hsp101. Plant Journal, 2005, 41, 722-731.	5.7	54
71	Developmental regulation of a proinsulin messenger RNA generated by intron retention. EMBO Reports, 2005, 6, 1182-1187.	4.5	44
72	Characterizing the function and structural organization of the 5' tRNA-like motif within the hepatitis C virus quasispecies. Nucleic Acids Research, 2005, 33, 1487-1502.	14.5	30

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73	Specific interference between two unrelated internal ribosome entry site elements impairs translation efficiency. FEBS Letters, 2005, 579, 6803-6808.	2.8	9
74	Internal Ribosome Entry Site Elements in Eukaryotic Genomes. Current Genomics, 2004, 5, 259-277.	1.6	3
75	Genome Organisation, Translation and Replication of Foot-and-Mouth Disease Virus RNA. , 2004, , 21-52.		2
76	Picornavirus IRES: Structure Function Relationship. Current Pharmaceutical Design, 2004, 10, 3757-3767.	1.9	38
77	Upstream AUGs in embryonic proinsulin mRNA control its low translation level. EMBO Journal, 2003, 22, 5582-5592.	7.8	47
78	Structural organization of a viral IRES depends on the integrity of the GNRA motif. Rna, 2003, 9, 1333-1344.	3.5	84
79	Stable expression of antisense RNAs targeted to the 5′ non-coding region confers heterotypic inhibition to foot-and-mouth disease virus infection. Journal of General Virology, 2003, 84, 393-402.	2.9	16
80	IRES-driven translation is stimulated separately by the FMDV 3'-NCR and poly(A) sequences. Nucleic Acids Research, 2002, 30, 4398-4405.	14.5	88
81	IRES elements: features of the RNA structure contributing to their activity. Biochimie, 2002, 84, 755-763.	2.6	23
82	Long-range RNA–RNA interactions between distant regions of the hepatitis C virus internal ribosome entry site element. Journal of General Virology, 2002, 83, 1113-1121.	2.9	31
83	IRES interaction with translation initiation factors: Functional characterization of novel RNA contacts with eIF3, eIF4B, and eIF4GII. Rna, 2001, 7, 1213-1226.	3.5	108
84	Deletion or substitution of the aphthovirus 3′ NCR abrogates infectivity and virus replication. Journal of General Virology, 2001, 82, 93-101.	2.9	72
85	Functional interactions in internal translation initiation directed by viral and cellular IRES elements. Journal of General Virology, 2001, 82, 973-984.	2.9	115
86	Interaction of the elF4G initiation factor with the aphthovirus IRES is essential for internal translation initiation in vivo. Rna, 2000, 6, 1380-1392.	3.5	121
87	Long-range RNA interactions between structural domains of the aphthovirus internal ribosome entry site (IRES). Rna, 1999, 5, 1374-1383.	3.5	69
88	Response To Retreatment With Interferon-α Plus Ribavirin in Chronic Hepatitis C Patients Is Independent of The Ns5a Gene Nucleotide Sequence. American Journal of Gastroenterology, 1999, 94, 2487-2495.	0.4	17
89	Internal ribosome entry site biology and its use in expression vectors. Current Opinion in Biotechnology, 1999, 10, 458-464.	6.6	181
90	Involvement of the Aphthovirus RNA Region Located between the Two Functional AUGs in Start Codon Selection. Virology, 1999, 255, 324-336.	2.4	71

4

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91	Internal initiation of translation efficiency in different hepatitis C genotypes isolated from interferon treated patients. Archives of Virology, 1999, 144, 215-229.	2.1	29
92	Heterotypic inhibition of foot-and-mouth disease virus infection by combinations of RNA transcripts corresponding to the $5\hat{a}\in^2$ and $3\hat{a}\in^2$ regions. Antiviral Research, 1999, 44, 133-141.	4.1	13
93	Response to retreatment with interferon-α plus ribavirin in chronic hepatitis C patients is independent of the NS5A gene nucleotide sequence. American Journal of Gastroenterology, 1999, 94, 2487-2495.	0.4	20
94	Parameters influencing translational efficiency in aphthovirus IRES-based bicistronic expression vectors. Gene, 1998, 217, 51-56.	2.2	27
95	Conserved structural motifs located in distal loops of aphthovirus internal ribosome entry site domain 3 are required for internal initiation of translation. Journal of Virology, 1997, 71, 4171-4175.	3.4	121
96	Molecular evolution of aphthoviruses. Virus Genes, 1995, 11, 197-207.	1.6	37
97	Effect of Expression of the Aphthovirus Protease 3C on Viral Infection and Gene Expression. Virology, 1995, 212, 111-120.	2.4	16
98	Picornavirus Variation. , 1993, , 255-281.		1
99	Primer design for specific diagnosis by PCR of highly variable RNA viruses: Typing of foot-and-mouth disease virus. Virology, 1992, 189, 363-367.	2.4	60
100	3D gene of foot-and-mouth disease virus. Journal of Molecular Biology, 1988, 204, 771-776.	4.2	27
101	Cloning and molecular characterization of a telomeric sequence from a temperature-induced Balbiani ring. Chromosoma, 1985, 92, 108-115.	2.2	53
102	The quasispecies (extremely heterogeneous) nature of viral RNA genome populations: biological relevance — a review. Gene, 1985, 40, 1-8.	2.2	484
103	Sequence of the viral replicase gene from foot-and-mouth disease virus C1-Santa Pau (C-S8). Gene, 1985, 35, 55-61.	2.2	42
104	Analysis of theilv-linked genes that determine the morphology ofEscherichia coli Cells. Current Microbiology, 1983, 8, 177-182.	2.2	1
105	Translation and Protein Processing. , 0, , 141-162.		9

106 Translation and Protein Processing. , 0, , 141-161.