

# Harald Janovjak

## List of Publications by Year in descending order

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Version: 2024-02-01

71  
papers

3,738  
citations

172457

29  
h-index

128289

60  
g-index

80  
all docs

80  
docs citations

80  
times ranked

4568  
citing authors

#	ARTICLE	IF	CITATIONS
1	Optogenetic neuroregeneration. <i>Neural Regeneration Research</i> , 2022, 17, 1468.	3.0	1
2	Light-activated receptor tyrosine kinases: Designs and applications. <i>Current Opinion in Pharmacology</i> , 2022, 63, 102197.	3.5	3
3	Structure-guided optimization of light-activated chimeric G-protein-coupled receptors. <i>Structure</i> , 2022, 30, 1075-1087.e4.	3.3	9
4	Optogenetic delivery of trophic signals in a genetic model of Parkinson's disease. <i>PLoS Genetics</i> , 2021, 17, e1009479.	3.5	11
5	Formation of Kiss1R/GPER Heterocomplexes Negatively Regulates Kiss1R-mediated Signalling through Limiting Receptor Cell Surface Expression. <i>Journal of Molecular Biology</i> , 2021, 433, 166843.	4.2	4
6	Microbial methionine transporters and biotechnological applications. <i>Applied Microbiology and Biotechnology</i> , 2021, 105, 3919-3929.	3.6	9
7	A Light-Oxygen-Voltage Receptor Integrates Light and Temperature. <i>Journal of Molecular Biology</i> , 2021, 433, 167107.	4.2	20
8	Acute and chronic effects of a light-activated FGF receptor in keratinocytes in vitro and in mice. <i>Life Science Alliance</i> , 2021, 4, e202101100.	2.8	5
9	A Rationally and Computationally Designed Fluorescent Biosensor for $\alpha$ -Serine. <i>ACS Sensors</i> , 2021, 6, 4193-4205.	7.8	8
10	LTP Induction Boosts Glutamate Spillover by Driving Withdrawal of Perisynaptic Astroglia. <i>Neuron</i> , 2020, 108, 919-936.e11.	8.1	159
11	Design and Application of Light-Regulated Receptor Tyrosine Kinases. <i>Methods in Molecular Biology</i> , 2020, 2173, 233-246.	0.9	4
12	All-Optical Miniaturized Co-culture Assay of Voltage-Gated Ca <sup>2+</sup> Channels. <i>Methods in Molecular Biology</i> , 2020, 2173, 247-260.	0.9	1
13	Optogenetic control of excitatory post-synaptic differentiation through neuroligin-1 tyrosine phosphorylation. <i>ELife</i> , 2020, 9, .	6.0	15
14	Editorial overview: Synthetic sensors and signals – new tools for a new trade. <i>Current Opinion in Structural Biology</i> , 2019, 57, iii-v.	5.7	0
15	Light-activated chimeric GPCRs: limitations and opportunities. <i>Current Opinion in Structural Biology</i> , 2019, 57, 196-203.	5.7	28
16	Engineering Strategy and Vector Library for the Rapid Generation of Modular Light-Controlled Protein-Protein Interactions. <i>Journal of Molecular Biology</i> , 2019, 431, 3046-3055.	4.2	19
17	Isolation of synaptic vesicles from genetically engineered cultured neurons. <i>Journal of Neuroscience Methods</i> , 2019, 312, 114-121.	2.5	1
18	Light-activated Frizzled7 reveals a permissive role of non-canonical wnt signaling in mesendoderm cell migration. <i>ELife</i> , 2019, 8, .	6.0	32

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19	Optical functionalization of human Class A orphan G-protein-coupled receptors. <i>Nature Communications</i> , 2018, 9, 1950.	12.8	46
20	Monitoring hippocampal glycine with the computationally designed optical sensor GlyFS. <i>Nature Chemical Biology</i> , 2018, 14, 861-869.	8.0	60
21	Optogenetic methods in drug screening: technologies and applications. <i>Current Opinion in Biotechnology</i> , 2017, 48, 8-14.	6.6	22
22	Gr�nlicht�induzierte Rezeptorinaktivierung durch Cobalamin�bindende Dom�nen. <i>Angewandte Chemie</i> , 2017, 129, 4679-4682.	2.0	5
23	P3.03-006 Optical Control of Growth Factor Receptors to Advance Signal Transduction Research and Drug Screening. <i>Journal of Thoracic Oncology</i> , 2017, 12, S1346-S1347.	1.1	0
24	Ancestral Protein Reconstruction and Circular Permutation for Improving the Stability and Dynamic Range of FRET Sensors. <i>Methods in Molecular Biology</i> , 2017, 1596, 71-87.	0.9	9
25	Green�Light�Induced Inactivation of Receptor Signaling Using Cobalamin�Binding Domains. <i>Angewandte Chemie - International Edition</i> , 2017, 56, 4608-4611.	13.8	85
26	Method for Developing Optical Sensors Using a Synthetic Dye-Fluorescent Protein FRET Pair and Computational Modeling and Assessment. <i>Methods in Molecular Biology</i> , 2017, 1596, 89-99.	0.9	2
27	Eine Phytochrom�Sensordom�ne erm�glicht eine Rezeptoraktivierung durch rotes Licht. <i>Angewandte Chemie</i> , 2016, 128, 6447-6450.	2.0	7
28	A Phytochrome Sensory Domain Permits Receptor Activation by Red Light. <i>Angewandte Chemie - International Edition</i> , 2016, 55, 6339-6342.	13.8	72
29	Optogenetic Control of Nodal Signaling Reveals a Temporal Pattern of Nodal Signaling Regulating Cell Fate Specification during Gastrulation. <i>Cell Reports</i> , 2016, 16, 866-877.	6.4	101
30	Rangefinder: A Semisynthetic FRET Sensor Design Algorithm. <i>ACS Sensors</i> , 2016, 1, 1286-1290.	7.8	11
31	Light at the End of the Protein: Crystal Structure of a C-Terminal Light-Sensing Domain. <i>Structure</i> , 2016, 24, 213-215.	3.3	1
32	Construction of a robust and sensitive arginine biosensor through ancestral protein reconstruction. <i>Protein Science</i> , 2015, 24, 1412-1422.	7.6	60
33	Quantification of riboflavin, flavin mononucleotide, and flavin adenine dinucleotide in mammalian model cells by CE with LED�induced fluorescence detection. <i>Electrophoresis</i> , 2015, 36, 518-525.	2.4	47
34	Light-assisted small-molecule screening against protein kinases. <i>Nature Chemical Biology</i> , 2015, 11, 952-954.	8.0	42
35	Flipping the Photoswitch: Ion Channels Under Light Control. <i>Advances in Experimental Medicine and Biology</i> , 2015, 869, 101-117.	1.6	12
36	The optogenetic promise for oncology: Episode I. <i>Molecular and Cellular Oncology</i> , 2014, 1, e964045.	0.7	5

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37	Spatio-temporally precise activation of engineered receptor tyrosine kinases by light. <i>EMBO Journal</i> , 2014, 33, 1713-1726.	7.8	226
38	Optical Control of Ligand-Gated Ion Channels. <i>Methods in Molecular Biology</i> , 2013, 998, 417-435.	0.9	3
39	Optical control of metabotropic glutamate receptors. <i>Nature Neuroscience</i> , 2013, 16, 507-516.	14.8	192
40	Optical Control of Metabotropic Glutamate Receptors for Probing of G Protein Signaling and Receptor Activation Mechanism. <i>Biophysical Journal</i> , 2012, 102, 517a.	0.5	0
41	Design and Application of a Light-Activated Metabotropic Glutamate Receptor for Optical Control of Intracellular Signaling Pathways. <i>Biophysical Journal</i> , 2011, 100, 177a.	0.5	0
42	A modern ionotropic glutamate receptor with a K <sup>+</sup> selectivity signature sequence. <i>Nature Communications</i> , 2011, 2, 232.	12.8	31
43	Pharmacology of ionotropic glutamate receptors: A structural perspective. <i>Bioorganic and Medicinal Chemistry</i> , 2010, 18, 7759-7772.	3.0	70
44	A light-gated, potassium-selective glutamate receptor for the optical inhibition of neuronal firing. <i>Nature Neuroscience</i> , 2010, 13, 1027-1032.	14.8	124
45	A Light-Gated, Potassium-Selective Glutamate Receptor for the Optical Inhibition of Neuronal Firing. <i>Biophysical Journal</i> , 2010, 98, 223a.	0.5	0
46	Periodic Forces Trigger a Complex Mechanical Response in Ubiquitin. <i>Journal of Molecular Biology</i> , 2009, 390, 443-456.	4.2	11
47	The Anisotropic Response of Ubiquitin Unfolded by Periodic Forces. <i>Biophysical Journal</i> , 2009, 96, 217a-218a.	0.5	0
48	Design Of A Potassium Selective, Light-gated Glutamate Receptor. <i>Biophysical Journal</i> , 2009, 96, 489a.	0.5	0
49	From Valleys to Ridges: Exploring the Dynamic Energy Landscape of Single Membrane Proteins. <i>ChemPhysChem</i> , 2008, 9, 954-966.	2.1	43
50	Fully automated single-molecule force spectroscopy for screening applications. <i>Nanotechnology</i> , 2008, 19, 384020.	2.6	32
51	Single-Molecule Microscopy and Force Spectroscopy of Membrane Proteins. <i>Springer Series in Biophysics</i> , 2008, , 279-311.	0.4	0
52	Digital force-feedback for protein unfolding experiments using atomic force microscopy. <i>Nanotechnology</i> , 2007, 18, 044022.	2.6	10
53	Deciphering Molecular Interactions of Native Membrane Proteins by Single-Molecule Force Spectroscopy. <i>Annual Review of Biophysics and Biomolecular Structure</i> , 2007, 36, 233-260.	18.3	124
54	Transmembrane Helices Have Rough Energy Surfaces. <i>Journal of the American Chemical Society</i> , 2007, 129, 246-247.	13.7	50

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55	Free Energy of Membrane Protein Unfolding Derived from Single-Molecule Force Measurements. <i>Biophysical Journal</i> , 2007, 93, 930-937.	0.5	45
56	Pulling single bacteriorhodopsin out of a membrane: Comparison of simulation and experiment. <i>Biochimica Et Biophysica Acta - Biomembranes</i> , 2006, 1758, 537-544.	2.6	24
57	Observing Folding Pathways and Kinetics of a Single Sodium-proton Antiporter from <i>Escherichia coli</i> . <i>Journal of Molecular Biology</i> , 2006, 355, 2-8.	4.2	48
58	Bacteriorhodopsin Folds into the Membrane against an External Force. <i>Journal of Molecular Biology</i> , 2006, 357, 644-654.	4.2	93
59	Imaging and detecting molecular interactions of single transmembrane proteins. <i>Neurobiology of Aging</i> , 2006, 27, 546-561.	3.1	38
60	Direct measurement of single-molecule visco-elasticity in atomic force microscope force-extension experiments. <i>European Biophysics Journal</i> , 2006, 35, 287-292.	2.2	24
61	Automated alignment and pattern recognition of single-molecule force spectroscopy data. <i>Journal of Microscopy</i> , 2005, 218, 125-132.	1.8	33
62	Hydrodynamic effects in fast AFM single-molecule force measurements. <i>European Biophysics Journal</i> , 2005, 34, 91-96.	2.2	111
63	Molecular Force Modulation Spectroscopy Revealing the Dynamic Response of Single Bacteriorhodopsins. <i>Biophysical Journal</i> , 2005, 88, 1423-1431.	0.5	69
64	Complex Stability of Single Proteins Explored by Forced Unfolding Experiments. <i>Biophysical Journal</i> , 2005, 88, L37-L39.	0.5	5
65	Probing the Energy Landscape of the Membrane Protein Bacteriorhodopsin. <i>Structure</i> , 2004, 12, 871-879.	3.3	80
66	Controlled Unfolding and Refolding of a Single Sodium-proton Antiporter using Atomic Force Microscopy. <i>Journal of Molecular Biology</i> , 2004, 340, 1143-1152.	4.2	99
67	Unfolding pathways of native bacteriorhodopsin depend on temperature. <i>EMBO Journal</i> , 2003, 22, 5220-5229.	7.8	111
68	Folding, Structure and Function of Biological Nanomachines Examined by AFM. <i>AIP Conference Proceedings</i> , 2003, , .	0.4	1
69	Cellular dynamics observed at sub-nanometer resolution using atomic force microscopy. <i>Microscopy and Microanalysis</i> , 2002, 8, 892-893.	0.4	0
70	Observing structure, function and assembly of single proteins by AFM. <i>Progress in Biophysics and Molecular Biology</i> , 2002, 79, 1-43.	2.9	155
71	Processing of gene expression data generated by quantitative real-time RT-PCR. <i>BioTechniques</i> , 2002, 32, 1372-4, 1376, 1378-9.	1.8	964