

Martin Jinek

List of Publications by Year in descending order

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82
papers

26,619
citations

57631

44
h-index

79541

73
g-index

106
all docs

106
docs citations

106
times ranked

26259
citing authors

#	ARTICLE	IF	CITATIONS
1	A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity. <i>Science</i> , 2012, 337, 816-821.	6.0	12,811
2	RNA-programmed genome editing in human cells. <i>ELife</i> , 2013, 2, e00471.	2.8	1,830
3	DNA interrogation by the CRISPR RNA-guided endonuclease Cas9. <i>Nature</i> , 2014, 507, 62-67.	13.7	1,573
4	Structural basis of PAM-dependent target DNA recognition by the Cas9 endonuclease. <i>Nature</i> , 2014, 513, 569-573.	13.7	1,075
5	Structures of Cas9 Endonucleases Reveal RNA-Mediated Conformational Activation. <i>Science</i> , 2014, 343, 1247997.	6.0	938
6	A three-dimensional view of the molecular machinery of RNA interference. <i>Nature</i> , 2009, 457, 405-412.	13.7	651
7	Sequence- and Structure-Specific RNA Processing by a CRISPR Endonuclease. <i>Science</i> , 2010, 329, 1355-1358.	6.0	599
8	A prudent path forward for genomic engineering and germline gene modification. <i>Science</i> , 2015, 348, 36-38.	6.0	541
9	Structural Biology of Nucleocytoplasmic Transport. <i>Annual Review of Biochemistry</i> , 2007, 76, 647-671.	5.0	458
10	Structural Basis for Guide RNA Processing and Seed-Dependent DNA Targeting by CRISPR-Cas12a. <i>Molecular Cell</i> , 2017, 66, 221-233.e4.	4.5	408
11	Structural insights into the molecular mechanism of the m6A writer complex. <i>ELife</i> , 2016, 5, .	2.8	386
12	Type III CRISPR-Cas systems produce cyclic oligoadenylate second messengers. <i>Nature</i> , 2017, 548, 543-548.	13.7	377
13	Mammalian miRNA RISC Recruits CAF1 and PABP to Affect PABP-Dependent Deadenylation. <i>Molecular Cell</i> , 2009, 35, 868-880.	4.5	331
14	Mechanistic Insights into the cis- and trans-Acting DNase Activities of Cas12a. <i>Molecular Cell</i> , 2019, 73, 589-600.e4.	4.5	298
15	The superhelical TPR-repeat domain of O-linked GlcNAc transferase exhibits structural similarities to importin β . <i>Nature Structural and Molecular Biology</i> , 2004, 11, 1001-1007.	3.6	263
16	Maximizing mutagenesis with solubilized CRISPR-Cas9 ribonucleoprotein complexes.. <i>Development (Cambridge)</i> , 2016, 143, 2025-37.	1.2	244
17	Structural Basis for DNase Activity of a Conserved Protein Implicated in CRISPR-Mediated Genome Defense. <i>Structure</i> , 2009, 17, 904-912.	1.6	228
18	In vivo adenine base editing of PCSK9 in macaques reduces LDL cholesterol levels. <i>Nature Biotechnology</i> , 2021, 39, 949-957.	9.4	196

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19	An RNA-induced conformational change required for CRISPR RNA cleavage by the endoribonuclease Cse3. <i>Nature Structural and Molecular Biology</i> , 2011, 18, 680-687.	3.6	166
20	Cas9 versus Cas12a/Cpf1: Structure–function comparisons and implications for genome editing. <i>Wiley Interdisciplinary Reviews RNA</i> , 2018, 9, e1481.	3.2	164
21	Structural Plasticity of PAM Recognition by Engineered Variants of the RNA-Guided Endonuclease Cas9. <i>Molecular Cell</i> , 2016, 61, 895-902.	4.5	161
22	Coupled 5' Nucleotide Recognition and Processivity in Xrn1-Mediated mRNA Decay. <i>Molecular Cell</i> , 2011, 41, 600-608.	4.5	155
23	CrisprVariants charts the mutation spectrum of genome engineering experiments. <i>Nature Biotechnology</i> , 2016, 34, 701-702.	9.4	149
24	CRISPR-Cas9 conformational activation as elucidated from enhanced molecular simulations. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2017, 114, 7260-7265.	3.3	133
25	Structural basis for the endoribonuclease activity of the type III-A CRISPR-associated protein Csm6. <i>Rna</i> , 2016, 22, 318-329.	1.6	128
26	Covalent linkage of the DNA repair template to the CRISPR-Cas9 nuclease enhances homology-directed repair. <i>ELife</i> , 2018, 7, .	2.8	127
27	DNA-guided DNA cleavage at moderate temperatures by <i>Clostridium butyricum</i> Argonaute. <i>Nucleic Acids Research</i> , 2019, 47, 5809-5821.	6.5	115
28	Structures of the tRNA export factor in the nuclear and cytosolic states. <i>Nature</i> , 2009, 461, 60-65.	13.7	108
29	Protospacer Adjacent Motif-Induced Allostery Activates CRISPR-Cas9. <i>Journal of the American Chemical Society</i> , 2017, 139, 16028-16031.	6.6	104
30	Striking Plasticity of CRISPR-Cas9 and Key Role of Non-target DNA, as Revealed by Molecular Simulations. <i>ACS Central Science</i> , 2016, 2, 756-763.	5.3	103
31	Deciphering Off-Target Effects in CRISPR-Cas9 through Accelerated Molecular Dynamics. <i>ACS Central Science</i> , 2019, 5, 651-662.	5.3	99
32	In Vitro Enzymology of Cas9. <i>Methods in Enzymology</i> , 2014, 546, 1-20.	0.4	97
33	Structural basis of AAUAAA polyadenylation signal recognition by the human CPSF complex. <i>Nature Structural and Molecular Biology</i> , 2018, 25, 135-138.	3.6	96
34	Structural insights into the human GW182-PABC interaction in microRNA-mediated deadenylation. <i>Nature Structural and Molecular Biology</i> , 2010, 17, 238-240.	3.6	92
35	An internal promoter underlies the difference in disease severity between N- and C-terminal truncation mutations of Titin in zebrafish. <i>ELife</i> , 2015, 4, e09406.	2.8	83
36	Mechanistic insights into mRNA 3'-end processing. <i>Current Opinion in Structural Biology</i> , 2019, 59, 143-150.	2.6	83

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37	Key role of the REC lobe during CRISPRâ€‘Cas9 activation by â€‘sensingâ€™™, â€‘regulatingâ€™™, and â€‘lockingâ€™™ the catalytic HNH domain. Quarterly Reviews of Biophysics, 2018, 51, .	2.4	79
38	Hakai is required for stabilization of core components of the m6A mRNA methylation machinery. Nature Communications, 2021, 12, 3778.	5.8	77
39	Structural insights into the assembly and polyA signal recognition mechanism of the human CPSF complex. ELife, 2017, 6, .	2.8	71
40	Evolution of CRISPR RNA recognition and processing by Cas6 endonucleases. Nucleic Acids Research, 2014, 42, 1341-1353.	6.5	68
41	Activation and self-inactivation mechanisms of the cyclic oligoadenylate-dependent CRISPR ribonuclease Csm6. Nature Communications, 2020, 11, 1596.	5.8	67
42	Molecular architectures and mechanisms of Class 2 CRISPR-associated nucleases. Current Opinion in Structural Biology, 2017, 47, 157-166.	2.6	65
43	Catalytic Mechanism of Non-Target DNA Cleavage in CRISPR-Cas9 Revealed by <i>Ab Initio</i> Molecular Dynamics. ACS Catalysis, 2020, 10, 13596-13605.	5.5	63
44	Bacteriophage DNA glucosylation impairs target DNA binding by type I and II but not by type V CRISPRâ€‘Cas effector complexes. Nucleic Acids Research, 2018, 46, 873-885.	6.5	57
45	Molecular architecture of <i>LSM</i> 14 interactions involved in the assembly of <i>mRNA</i> silencing complexes. EMBO Journal, 2018, 37, .	3.5	51
46	Molecular basis for cytoplasmic <i>RNA</i> surveillance by uridylationâ€‘triggered decay in <i>Drosophila</i> . EMBO Journal, 2016, 35, 2417-2434.	3.5	50
47	Molecular Dynamics Reveals a DNA-Induced Dynamic Switch Triggering Activation of CRISPR-Cas12a. Journal of Chemical Information and Modeling, 2020, 60, 6427-6437.	2.5	43
48	Data-collection strategy for challenging native SAD phasing. Acta Crystallographica Section D: Structural Biology, 2016, 72, 421-429.	1.1	42
49	Target site selection and remodelling by type V CRISPR-transposon systems. Nature, 2021, 599, 497-502.	13.7	42
50	Structural insights into the Notch-modifying glycosyltransferase Fringe. Nature Structural and Molecular Biology, 2006, 13, 945-946.	3.6	35
51	Molecular mechanism of the RNA helicase DHX37 and its activation by UTP14A in ribosome biogenesis. Rna, 2019, 25, 685-701.	1.6	33
52	Structural and Biochemical Studies of a Fluoroacetyl-CoA-Specific Thioesterase Reveal a Molecular Basis for Fluorine Selectivity. Biochemistry, 2010, 49, 9269-9279.	1.2	31
53	The C-terminal region of Ge-1 presents conserved structural features required for P-body localization. Rna, 2008, 14, 1991-1998.	1.6	30
54	Structural mimicry in transcription regulation of human RNA polymerase II by the DNA helicase RECQL5. Nature Structural and Molecular Biology, 2013, 20, 892-899.	3.6	27

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55	Conformational control of Cas9 by CRISPR hybrid RNA-DNA guides mitigates off-target activity in TÀcells. <i>Molecular Cell</i> , 2021, 81, 3637-3649.e5.	4.5	27
56	In Vitro Reconstitution and Crystallization of Cas9 Endonuclease Bound to a Guide RNA and a DNA Target. <i>Methods in Enzymology</i> , 2015, 558, 515-537.	0.4	23
57	ANGEL2 is a member of the CCR4 family of deadenylases with 2â€²,3â€²-cyclic phosphatase activity. <i>Science</i> , 2020, 369, 524-530.	6.0	23
58	Human MARF1 is an endoribonuclease that interacts with the DCP1:2 decapping complex and degrades target mRNAs. <i>Nucleic Acids Research</i> , 2018, 46, 12008-12021.	6.5	22
59	Molecular architecture of the human tRNA ligase complex. <i>ELife</i> , 2021, 10, .	2.8	22
60	The oxidoreductase PYROXD1 uses NAD(P)+ as an antioxidant to sustain tRNA ligase activity in pre-tRNA splicing and unfolded protein response. <i>Molecular Cell</i> , 2021, 81, 2520-2532.e16.	4.5	21
61	Heterologous Expression and Purification of the CRISPR-Cas12a/Cpf1 Protein. <i>Bio-protocol</i> , 2018, 8, e2842.	0.2	21
62	Introducing gene deletions by mouse zygote electroporation of Cas12a/Cpf1. <i>Transgenic Research</i> , 2019, 28, 525-535.	1.3	20
63	Preparation and electroporation of Cas12a/Cpf1-guide RNA complexes for introducing large gene deletions in mouse embryonic stem cells. <i>Methods in Enzymology</i> , 2019, 616, 241-263.	0.4	16
64	Crystal structure of the C-terminal 2â€²,5â€²-phosphodiesterase domain of group a rotavirus protein VP3. <i>Proteins: Structure, Function and Bioinformatics</i> , 2015, 83, 997-1002.	1.5	14
65	Structural basis for acceptor RNA substrate selectivity of the 3â€² terminal uridylyl transferase Tailor. <i>Nucleic Acids Research</i> , 2019, 47, 1030-1042.	6.5	13
66	In vitro Generation of CRISPR-Cas9 Complexes with Covalently Bound Repair Templates for Genome Editing in Mammalian Cells. <i>Bio-protocol</i> , 2019, 9, .	0.2	13
67	Use of RNA Tertiary Interaction Modules for the Crystallisation of the Spliceosomal snRNP Core Domain. <i>Journal of Molecular Biology</i> , 2010, 402, 154-164.	2.0	11
68	CRISPR-Directed Therapeutic Correction at the NCF1 Locus Is Challenged by Frequent Incidence of Chromosomal Deletions. <i>Molecular Therapy - Methods and Clinical Development</i> , 2020, 17, 936-943.	1.8	8
69	Multiplexed Single-Molecule Experiments Reveal Nucleosome Invasion Dynamics of the Cas9 Genome Editor. <i>Journal of the American Chemical Society</i> , 2021, 143, 16313-16319.	6.6	6
70	Specialized Weaponry: How a Type III-A CRISPR-Cas System Excels at Combating Phages. <i>Cell Host and Microbe</i> , 2017, 22, 258-259.	5.1	5
71	Two-Metal Ion Mechanism of DNA Cleavage in CRISPR-Cas9. <i>Biophysical Journal</i> , 2020, 118, 64a.	0.2	2
72	The CRISPR-RNA World: An Interview with Martin JÅnek. <i>CRISPR Journal</i> , 2020, 3, 68-72.	1.4	2

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73	The Oxidoreductase PYROXD1 Utilizes NAD(P)+ As an Antioxidant to Sustain tRNA Ligase Activity in Pre-tRNA Splicing and Unfolded Protein Response. SSRN Electronic Journal, 0, , .	0.4	1
74	Eukaryotic expression, purification, crystallization and preliminary X-ray analysis of murine Manic Fringe. Acta Crystallographica Section F: Structural Biology Communications, 2006, 62, 774-777.	0.7	0
75	CRISPR-Cas9: Computational Insights Toward Improved Genome Editing. Biophysical Journal, 2017, 112, 72a.	0.2	0
76	Cover Image, Volume 9, Issue 5. Wiley Interdisciplinary Reviews RNA, 2018, 9, e1505.	3.2	0
77	A PAM-Induced Signalling Activates the Communication between HNH and RUVF in CRISPR-Cas9. Biophysical Journal, 2018, 114, 250a.	0.2	0
78	Editorial overview: Proteinâ€™nucleic acid interactions â€™ cryo-EM, what else?. Current Opinion in Structural Biology, 2019, 59, vi-viii.	2.6	0
79	Uncut but Primed for Change. CRISPR Journal, 2019, 2, 352-354.	1.4	0
80	Critical Role of Conserved Histidine Residues in Genome Editing and Recombination. Biophysical Journal, 2021, 120, 137a-138a.	0.2	0
81	Cooperative Dynamics of REC-Nuc Lobes Prime Cas12a for DNA Processing. Biophysical Journal, 2021, 120, 16a-17a.	0.2	0
82	Multi-microsecond molecular dynamics unveils the mechanism of DNA traversal within CRISPR-Cas12a. Biophysical Journal, 2022, 121, 322a.	0.2	0