

Alexis Vallée-Bélisle

List of Publications by Year in descending order

Source: <https://exaly.com/author-pdf/4793919/publications.pdf>

Version: 2024-02-01

50
papers

4,217
citations

101543

36
h-index

182427

51
g-index

54
all docs

54
docs citations

54
times ranked

4746
citing authors

#	ARTICLE	IF	CITATIONS
1	Monitoring protein conformational changes using fluorescent nanoantennas. <i>Nature Methods</i> , 2022, 19, 71-80.	19.0	17
2	Silver oxide model surface improves computational simulation of surface-enhanced Raman spectroscopy on silver nanoparticles. <i>Physical Chemistry Chemical Physics</i> , 2021, 23, 15480-15484.	2.8	1
3	Optimizing the Specificity Window of Biomolecular Receptors Using Structure-Switching and Allostery. <i>ACS Sensors</i> , 2020, 5, 1937-1942.	7.8	14
4	Peptide-Mediated Electrochemical Steric Hindrance Assay for One-Step Detection of HIV Antibodies. <i>Analytical Chemistry</i> , 2019, 91, 4943-4947.	6.5	35
5	Programmable DNA switches and their applications. <i>Nanoscale</i> , 2018, 10, 4607-4641.	5.6	101
6	Engineering Biosensors with Dual Programmable Dynamic Ranges. <i>Analytical Chemistry</i> , 2018, 90, 1506-1510.	6.5	19
7	Aptamer-based liposomes improve specific drug loading and release. <i>Journal of Controlled Release</i> , 2017, 251, 82-91.	9.9	46
8	Steric Hindrance Assay for Secreted Factors in Stem Cell Culture. <i>ACS Sensors</i> , 2017, 2, 495-500.	7.8	14
9	Electrochemical DNA-Based Immunoassay That Employs Steric Hindrance To Detect Small Molecules Directly in Whole Blood. <i>ACS Sensors</i> , 2017, 2, 718-723.	7.8	45
10	Antibody-powered nucleic acid release using a DNA-based nanomachine. <i>Nature Communications</i> , 2017, 8, 15150.	12.8	108
11	A DNA Nanodevice That Loads and Releases a Cargo with Hemoglobin-Like Allosteric Control and Cooperativity. <i>Nano Letters</i> , 2017, 17, 3225-3230.	9.1	25
12	Biomolecular Steric Hindrance Effects Are Enhanced on Nanostructured Microelectrodes. <i>Analytical Chemistry</i> , 2017, 89, 9751-9757.	6.5	39
13	Determining the folding and binding free energy of DNA-based nanodevices and nanoswitches using urea titration curves. <i>Nucleic Acids Research</i> , 2017, 45, 7571-7580.	14.5	26
14	Programmable Quantitative DNA Nanothermometers. <i>Nano Letters</i> , 2016, 16, 3976-3981.	9.1	67
15	Using Nature's "Tricks" To Rationally Tune the Binding Properties of Biomolecular Receptors. <i>Accounts of Chemical Research</i> , 2016, 49, 1884-1892.	15.6	123
16	A Modular, DNA-Based Beacon for Single-Step Fluorescence Detection of Antibodies and Other Proteins. <i>Angewandte Chemie - International Edition</i> , 2015, 54, 13214-13218.	13.8	93
17	Electrochemical structure-switching sensing using nanoplasmonic devices. <i>Annalen Der Physik</i> , 2015, 527, 806-813.	2.4	4
18	Controlling Hybridization Chain Reactions with pH. <i>Nano Letters</i> , 2015, 15, 5539-5544.	9.1	49

#	ARTICLE	IF	CITATIONS
19	General Strategy to Introduce pH-Induced Allostery in DNA-Based Receptors to Achieve Controlled Release of Ligands. <i>Nano Letters</i> , 2015, 15, 4467-4471.	9.1	91
20	Electrochemical plasmonic sensing system for highly selective multiplexed detection of biomolecules based on redox nanoswitches. <i>Biosensors and Bioelectronics</i> , 2015, 71, 75-81.	10.1	26
21	A Highly Selective Electrochemical DNA-Based Sensor That Employs Steric Hindrance Effects to Detect Proteins Directly in Whole Blood. <i>Journal of the American Chemical Society</i> , 2015, 137, 15596-15599.	13.7	162
22	Enzyme-Operated DNA-Based Nanodevices. <i>Nano Letters</i> , 2015, 15, 8407-8411.	9.1	46
23	A comparison of the folding kinetics of a small, artificially selected DNA aptamer with those of equivalently simple naturally occurring proteins. <i>Protein Science</i> , 2014, 23, 56-66.	7.6	12
24	Intrinsic disorder as a generalizable strategy for the rational design of highly responsive, allosterically cooperative receptors. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2014, 111, 15048-15053.	7.1	69
25	Programmable pH-Triggered DNA Nanoswitches. <i>Journal of the American Chemical Society</i> , 2014, 136, 5836-5839.	13.7	296
26	Using the Population-Shift Mechanism to Rationally Introduce "Hill-type" Cooperativity into a Normally Non-Cooperative Receptor. <i>Angewandte Chemie - International Edition</i> , 2014, 53, 9471-9475.	13.8	41
27	Principles for the Rational Design of Allosterically Cooperative Biomolecular Receptors. <i>Biophysical Journal</i> , 2014, 106, 614a.	0.5	0
28	Thermodynamic Basis for Engineering High-Affinity, High-Specificity Binding-Induced DNA Clamp Nanoswitches. <i>ACS Nano</i> , 2013, 7, 10863-10869.	14.6	58
29	Allosterically Tunable, DNA-Based Switches Triggered by Heavy Metals. <i>Journal of the American Chemical Society</i> , 2013, 135, 13238-13241.	13.7	99
30	DNA biomolecular-electronic encoder and decoder devices constructed by multiplex biosensors. <i>NPG Asia Materials</i> , 2012, 4, e1-e1.	7.9	138
31	Engineering Biosensors with Extended, Narrowed, or Arbitrarily Edited Dynamic Range. <i>Journal of the American Chemical Society</i> , 2012, 134, 2876-2879.	13.7	135
32	Employing the Metabolic "Branch Point Effect" to Generate an All-or-None, Digital-like Response in Enzymatic Outputs and Enzyme-Based Sensors. <i>Analytical Chemistry</i> , 2012, 84, 1076-1082.	6.5	41
33	Entropic and Electrostatic Effects on the Folding Free Energy of a Surface-Attached Biomolecule: An Experimental and Theoretical Study. <i>Journal of the American Chemical Society</i> , 2012, 134, 2120-2126.	13.7	47
34	Rational Design of Allosteric Inhibitors and Activators Using the Population-Shift Model: In Vitro Validation and Application to an Artificial Biosensor. <i>Journal of the American Chemical Society</i> , 2012, 134, 15177-15180.	13.7	80
35	Using Distal-Site Mutations and Allosteric Inhibition To Tune, Extend, and Narrow the Useful Dynamic Range of Aptamer-Based Sensors. <i>Journal of the American Chemical Society</i> , 2012, 134, 20601-20604.	13.7	132
36	Bioelectrochemical Switches for the Quantitative Detection of Antibodies Directly in Whole Blood. <i>Journal of the American Chemical Society</i> , 2012, 134, 15197-15200.	13.7	103

#	ARTICLE	IF	CITATIONS
37	Quantification of Transcription Factor Binding in Cell Extracts Using an Electrochemical, Structure-Switching Biosensor. <i>Journal of the American Chemical Society</i> , 2012, 134, 3346-3348.	13.7	81
38	Re-engineering Electrochemical Biosensors To Narrow or Extend Their Useful Dynamic Range. <i>Angewandte Chemie - International Edition</i> , 2012, 51, 6717-6721.	13.8	80
39	Visualizing transient protein-folding intermediates by tryptophan-scanning mutagenesis. <i>Nature Structural and Molecular Biology</i> , 2012, 19, 731-736.	8.2	48
40	Transcription Factor Beacons for the Quantitative Detection of DNA Binding Activity. <i>Journal of the American Chemical Society</i> , 2011, 133, 13836-13839.	13.7	79
41	High-Precision, In Vitro Validation of the Sequestration Mechanism for Generating Ultrasensitive Dose-Response Curves in Regulatory Networks. <i>PLoS Computational Biology</i> , 2011, 7, e1002171.	3.2	44
42	Structure-switching biosensors: inspired by Nature. <i>Current Opinion in Structural Biology</i> , 2010, 20, 518-526.	5.7	163
43	Colorimetric detection of DNA, small molecules, proteins, and ions using unmodified gold nanoparticles and conjugated polyelectrolytes. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2010, 107, 10837-10841.	7.1	505
44	Label-Free, Dual-Analyte Electrochemical Biosensors: A New Class of Molecular-Electronic Logic Gates. <i>Journal of the American Chemical Society</i> , 2010, 132, 8557-8559.	13.7	117
45	On the Binding of Cationic, Water-Soluble Conjugated Polymers to DNA: Electrostatic and Hydrophobic Interactions. <i>Journal of the American Chemical Society</i> , 2010, 132, 1252-1254.	13.7	82
46	Using Triplex-Forming Oligonucleotide Probes for the Reagentless, Electrochemical Detection of Double-Stranded DNA. <i>Analytical Chemistry</i> , 2010, 82, 9109-9115.	6.5	87
47	Thermodynamic basis for the optimization of binding-induced biomolecular switches and structure-switching biosensors. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 2009, 106, 13802-13807.	7.1	146
48	Multiple Tryptophan Probes Reveal that Ubiquitin Folds via a Late Misfolded Intermediate. <i>Journal of Molecular Biology</i> , 2007, 374, 791-805.	4.2	28
49	Protein folding: Defining a "standard" set of experimental conditions and a preliminary kinetic data set of two-state proteins. <i>Protein Science</i> , 2005, 14, 602-616.	7.6	207
50	[14] Detection of protein-protein interactions by protein fragment complementation strategies. <i>Methods in Enzymology</i> , 2000, 328, 208-230.	1.0	117