

Audrey Claing

List of Publications by Year in descending order

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34
papers

2,594
citations

304743

22
h-index

377865

34
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34
all docs

34
docs citations

34
times ranked

2969
citing authors

#	ARTICLE	IF	CITATIONS
1	Activation of the GTPase ARF6 regulates invasion of human vascular smooth muscle cells by stimulating MMP14 activity. <i>Scientific Reports</i> , 2022, 12, .	3.3	3
2	Discovery of a dual Ras and ARF6 inhibitor from a GPCR endocytosis screen. <i>Nature Communications</i> , 2021, 12, 4688.	12.8	7
3	Methods to Investigate the β -Arrestin-Mediated Control of ARF6 Activation to Regulate Trafficking and Actin Cytoskeleton Remodeling. <i>Methods in Molecular Biology</i> , 2019, 1957, 159-168.	0.9	3
4	ARF GTPases control phenotypic switching of vascular smooth muscle cells through the regulation of actin function and actin dependent gene expression. <i>Cellular Signalling</i> , 2018, 46, 64-75.	3.6	9
5	ARF6 protects sister chromatid cohesion to ensure the formation of stable kinetochore-microtubule attachments. <i>Journal of Cell Science</i> , 2018, 131, .	2.0	2
6	Arterial stiffness induced by carotid calcification leads to cerebral gliosis mediated by oxidative stress. <i>Journal of Hypertension</i> , 2018, 36, 286-298.	0.5	22
7	β -Arrestin-mediated Angiotensin II Signaling Controls the Activation of ARF6 Protein and Endocytosis in Migration of Vascular Smooth Muscle Cells. <i>Journal of Biological Chemistry</i> , 2016, 291, 3967-3981.	3.4	22
8	The GTPase ARF6 Controls ROS Production to Mediate Angiotensin II-Induced Vascular Smooth Muscle Cell Proliferation. <i>PLoS ONE</i> , 2016, 11, e0148097.	2.5	35
9	ADP-ribosylation factor 1 expression regulates epithelial-mesenchymal transition and predicts poor clinical outcome in triple-negative breast cancer. <i>Oncotarget</i> , 2016, 7, 15811-15827.	1.8	18
10	The small GTPase ADP-Ribosylation Factor 1 mediates the sensitivity of triple negative breast cancer cells to EGFR tyrosine kinase inhibitors. <i>Cancer Biology and Therapy</i> , 2015, 16, 1535-1547.	3.4	18
11	ARF1 regulates adhesion of MDA-MB-231 invasive breast cancer cells through formation of focal adhesions. <i>Cellular Signalling</i> , 2015, 27, 403-415.	3.6	40
12	ARF1 regulates the Rho/MLC pathway to control EGF-dependent breast cancer cell invasion. <i>Molecular Biology of the Cell</i> , 2014, 25, 17-29.	2.1	116
13	The Adaptor Proteins p66Shc and Grb2 Regulate the Activation of the GTPases ARF1 and ARF6 in Invasive Breast Cancer Cells. <i>Journal of Biological Chemistry</i> , 2014, 289, 5687-5703.	3.4	50
14	β -Arrestins. <i>Progress in Molecular Biology and Translational Science</i> , 2013, 118, 149-174.	1.7	11
15	ARF1 controls Rac1 signaling to regulate migration of MDA-MB-231 invasive breast cancer cells. <i>Cellular Signalling</i> , 2013, 25, 1813-1819.	3.6	36
16	Differential β -Arrestin-Dependent Conformational Signaling and Cellular Responses Revealed by Angiotensin Analogs. <i>Science Signaling</i> , 2012, 5, ra33.	3.6	140
17	Vascular Endothelial Growth Factor Receptor-2 Activates ADP-ribosylation Factor 1 to Promote Endothelial Nitric-oxide Synthase Activation and Nitric Oxide Release from Endothelial Cells. <i>Journal of Biological Chemistry</i> , 2010, 285, 24591-24599.	3.4	18
18	G protein-coupled receptors stimulation and the control of cell migration. <i>Cellular Signalling</i> , 2009, 21, 1045-1053.	3.6	143

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19	Endothelin-1 promotes migration of endothelial cells through the activation of ARF6 and the regulation of FAK activity. <i>Cellular Signalling</i> , 2008, 20, 2256-2265.	3.6	34
20	ADP-ribosylation Factor 1 Controls the Activation of the Phosphatidylinositol 3-Kinase Pathway to Regulate Epidermal Growth Factor-dependent Growth and Migration of Breast Cancer Cells. <i>Journal of Biological Chemistry</i> , 2008, 283, 36425-36434.	3.4	83
21	Endogenous ARF6 Interacts with Rac1 upon Angiotensin II Stimulation to Regulate Membrane Ruffling and Cell Migration. <i>Molecular Biology of the Cell</i> , 2007, 18, 501-511.	2.1	60
22	ARF6 regulates angiotensin II type 1 receptor endocytosis by controlling the recruitment of AP-2 and clathrin. <i>Cellular Signalling</i> , 2007, 19, 2370-2378.	3.6	34
23	ARF6 activation by G β q signaling: G β q forms molecular complexes with ARNO and ARF6. <i>Cellular Signalling</i> , 2006, 18, 1988-1994.	3.6	27
24	G Protein-coupled Receptor Endocytosis in ADP-ribosylation Factor 6-depleted Cells. <i>Journal of Biological Chemistry</i> , 2005, 280, 5598-5604.	3.4	70
25	Novel roles for arrestins in G protein-coupled receptor biology and drug discovery. <i>Current Opinion in Drug Discovery & Development</i> , 2005, 8, 585-9.	1.9	4
26	Regulation of G protein-coupled receptor endocytosis by ARF6 GTP-binding proteins. <i>Biochemistry and Cell Biology</i> , 2004, 82, 610-617.	2.0	22
27	Endocytosis of G protein-coupled receptors: roles of G protein-coupled receptor kinases and γ -arrestin proteins. <i>Progress in Neurobiology</i> , 2002, 66, 61-79.	5.7	493
28	Comparison of the contractile and calcium-increasing properties of platelet-activating factor and endothelin-1 in the rat mesenteric artery and vein. <i>British Journal of Pharmacology</i> , 2002, 135, 433-443.	5.4	26
29	β 2-Arrestin-mediated ADP-ribosylation Factor 6 Activation and β 2-Adrenergic Receptor Endocytosis. <i>Journal of Biological Chemistry</i> , 2001, 276, 42509-42513.	3.4	204
30	The GIT Family of ADP-ribosylation Factor GTPase-activating Proteins. <i>Journal of Biological Chemistry</i> , 2000, 275, 22373-22380.	3.4	125
31	The β 2-adrenergic receptor interacts with the Na ⁺ /H ⁺ -exchanger regulatory factor to control Na ⁺ /H ⁺ exchange. <i>Nature</i> , 1998, 392, 626-630.	27.8	566
32	Processing of proendothelin-1 by human furin convertase. <i>FEBS Letters</i> , 1995, 362, 276-280.	2.8	91
33	Role of α -type calcium channels in the response of the perfused arterial and venous mesenteric vasculature of the rat to platelet-activating factor. <i>British Journal of Pharmacology</i> , 1994, 112, 1202-1208.	5.4	22
34	Different pharmacological profiles of big β -endothelin-1 and big α -endothelin-1 <i>in vivo</i> and <i>in vitro</i> . <i>British Journal of Pharmacology</i> , 1991, 104, 440-444.	5.4	40