# VÃ-ctor De Lorenzo

List of Publications by Year in descending order

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217 papers	14,939 citations	<sup>16451</sup> 64 h-index	23533 111 g-index
237	237	237	10177
all docs	docs citations	times ranked	citing authors

#	Article	IF	CITATIONS
1	Standardization of regulatory nodes for engineering heterologous gene expression: a feasibility study. Microbial Biotechnology, 2022, 15, 2250-2265.	4.2	8
2	High-Efficiency Multi-site Genomic Editing (HEMSE) Made Easy. Methods in Molecular Biology, 2022, 2479, 37-52.	0.9	0
3	Hypermutation of specific genomic loci of <i>Pseudomonas putida</i> for continuous evolution of target genes. Microbial Biotechnology, 2022, 15, 2309-2323.	4.2	3
4	Environmental Galenics: large-scale fortification of extant microbiomes with engineered bioremediation agents. Philosophical Transactions of the Royal Society B: Biological Sciences, 2022, 377, .	4.0	13
5	For the sake of the Bioeconomy: define what a Synthetic Biology Chassis is!. New Biotechnology, 2021, 60, 44-51.	4.4	34
6	A Standardized Inverter Package Borne by Broad Host Range Plasmids for Genetic Circuit Design in Gram-Negative Bacteria. ACS Synthetic Biology, 2021, 10, 213-217.	3.8	9
7	Ribonucleases control distinct traits of <i>Pseudomonas putida</i> lifestyle. Environmental Microbiology, 2021, 23, 174-189.	3.8	5
8	Reconfiguration of metabolic fluxes in <i>Pseudomonas putida</i> as a response to sub-lethal oxidative stress. ISME Journal, 2021, 15, 1751-1766.	9.8	79
9	Low CyaA expression and antiâ€cooperative binding of cAMP to CRP frames the scope of the cognate regulon of Pseudomonas putida. Environmental Microbiology, 2021, 23, 1732-1749.	3.8	4
10	Subcellular Architecture of the <i>xyl</i> Gene Expression Flow of the TOL Catabolic Plasmid of Pseudomonas putida mt-2. MBio, 2021, 12, .	4.1	3
11	A Bifan Motif Shaped by ArsR1, ArsR2, and Their Cognate Promoters Frames Arsenic Tolerance of Pseudomonas putida. Frontiers in Microbiology, 2021, 12, 641440.	3.5	2
12	Refactoring the Conjugation Machinery of Promiscuous Plasmid RP4 into a Device for Conversion of Gram-Negative Isolates to Hfr Strains. ACS Synthetic Biology, 2021, 10, 690-697.	3.8	7
13	Identification of a selfâ€sufficient cytochrome P450 monooxygenase from <i>Cupriavidus pinatubonensis</i> JMP134 involved in 2â€hydroxyphenylacetic acid catabolism, via homogentisate pathway. Microbial Biotechnology, 2021, 14, 1944-1960.	4.2	7
14	Engineering Tropism of <i>Pseudomonas putida</i> toward Target Surfaces through Ectopic Display of Recombinant Nanobodies. ACS Synthetic Biology, 2021, 10, 2049-2059.	3.8	11
15	Automated design and implementation of a NOR gate in Pseudomonas putida. Synthetic Biology, 2021, 6, ysab024.	2.2	12
16	The faulty SOS response of Pseudomonas putida KT2440 stems from an inefficient RecA‣exA interplay. Environmental Microbiology, 2021, 23, 1608-1619.	3.8	0
17	Targetron-Assisted Delivery of Exogenous DNA Sequences into <i>Pseudomonas putida</i> through CRISPR-Aided Counterselection. ACS Synthetic Biology, 2021, 10, 2552-2565.	3.8	8
18	Mismatch repair hierarchy of <i>Pseudomonas putida</i> revealed by mutagenic ssDNA recombineering of the <i>pyrF</i> gene. Environmental Microbiology, 2020, 22, 45-58.	3.8	22

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19	SEVA 3.0: an update of the Standard European Vector Architecture for enabling portability of genetic constructs among diverse bacterial hosts. Nucleic Acids Research, 2020, 48, D1164-D1170.	14.5	82
20	Multiple-Site Diversification of Regulatory Sequences Enables Interspecies Operability of Genetic Devices. ACS Synthetic Biology, 2020, 9, 104-114.	3.8	15
21	An automated DIY framework for experimental evolution ofPseudomonas putida. Microbial Biotechnology, 2020, 14, 2679-2685.	4.2	5
22	SEVA 3.1: enabling interoperability of DNA assembly among the SEVA, BioBricks and Type IIS restriction enzyme standards. Microbial Biotechnology, 2020, 13, 1793-1806.	4.2	26
23	Naked Bacterium: Emerging Properties of a Surfome-Streamlined <i>Pseudomonas putida</i> Strain. ACS Synthetic Biology, 2020, 9, 2477-2492.	3.8	15
24	In vivo diversification of target genomic sites using processive base deaminase fusions blocked by dCas9. Nature Communications, 2020, 11, 6436.	12.8	47
25	The Wsp intermembrane complex mediates metabolic control of the swimâ€∎ttach decision of <i>Pseudomonas putida</i> . Environmental Microbiology, 2020, 22, 3535-3547.	3.8	13
26	High-Efficiency Multi-site Genomic Editing of Pseudomonas putida through Thermoinducible ssDNA Recombineering. IScience, 2020, 23, 100946.	4.1	32
27	<scp>ArsH</scp> protects <i>Pseudomonas putida</i> from oxidative damage caused by exposure to arsenic. Environmental Microbiology, 2020, 22, 2230-2242.	3.8	28
28	Multifunctional SEVA shuttle vectors for actinomycetes and Gramâ€negative bacteria. MicrobiologyOpen, 2020, 9, 1135-1149.	3.0	12
29	Environmental Performance of <i>Pseudomonas putida</i> with a Uracylated Genome. ChemBioChem, 2020, 21, 3255-3265.	2.6	3
30	Synthetic Biology for Terraformation Lessons from Mars, Earth, and the Microbiome. Life, 2020, 10, 14.	2.4	28
31	A Broad Host Range Plasmid-Based Roadmap for ssDNA-Based Recombineering in Gram-Negative Bacteria. Methods in Molecular Biology, 2020, 2075, 383-398.	0.9	11
32	<scp>CRISPR</scp> /Cas9â€enhanced ss <scp>DNA</scp> recombineering for <i>Pseudomonas putida</i> . Microbial Biotechnology, 2019, 12, 1076-1089.	4.2	31
33	Spatial organization of the gene expression hardware in <i>Pseudomonas putida</i> . Environmental Microbiology, 2019, 21, 1645-1658.	3.8	14
34	Genomic Responses of Pseudomonas putida to Aromatic Hydrocarbons. , 2019, , 1-15.		0
35	Functional implementation of a linear glycolysis for sugar catabolism in Pseudomonas putida. Metabolic Engineering, 2019, 54, 200-211.	7.0	56
36	Pseudomonas putida in the quest of programmable chemistry. Current Opinion in Biotechnology, 2019, 59, 111-121.	6.6	38

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37	Improved Thermotolerance of Genomeâ€Reduced <i>Pseudomonas putida</i> EM42 Enables Effective Functioning of the P <sub>L</sub> / <i>c</i> 1857 System. Biotechnology Journal, 2019, 14, e1800483.	3.5	27
38	Evolving metabolism of 2,4â€dinitrotoluene triggers SOSâ€independent diversification of host cells. Environmental Microbiology, 2019, 21, 314-326.	3.8	13
39	Genomic Responses of Pseudomonas putida to Aromatic Hydrocarbons. , 2019, , 287-301.		0
40	Biodegradation and Bioremediation: An Introduction. , 2019, , 1-20.		0
41	Assessing Carbon Source-Dependent Phenotypic Variability in Pseudomonas putida. Methods in Molecular Biology, 2018, 1745, 287-301.	0.9	4
42	The power of synthetic biology for bioproduction, remediation and pollution control. EMBO Reports, 2018, 19, .	4.5	83
43	CRISPR/Cas9â€Based Counterselection Boosts Recombineering Efficiency in <i>Pseudomonas putida</i> . Biotechnology Journal, 2018, 13, e1700161.	3.5	115
44	A standardized workflow for surveying recombinases expands bacterial genomeâ€editing capabilities. Microbial Biotechnology, 2018, 11, 176-188.	4.2	43
45	A Post-translational Metabolic Switch Enables Complete Decoupling of Bacterial Growth from Biopolymer Production in Engineered <i>Escherichia coli</i> . ACS Synthetic Biology, 2018, 7, 2686-2697.	3.8	58
46	The interplay of EIIA <sup>Ntr</sup> with Câ€source regulation of the <i>Pu</i> promoter of <i>Pseudomonas putida</i> mtâ€2. Environmental Microbiology, 2018, 20, 4555-4566.	3.8	3
47	The Metabolic Redox Regime of Pseudomonas putida Tunes Its Evolvability toward Novel Xenobiotic Substrates. MBio, 2018, 9, .	4.1	51
48	Pseudomonas putida as a functional chassis for industrial biocatalysis: From native biochemistry to trans-metabolism. Metabolic Engineering, 2018, 50, 142-155.	7.0	338
49	Refactoring the upper sugar metabolism of Pseudomonas putida for co-utilization of cellobiose, xylose, and glucose. Metabolic Engineering, 2018, 48, 94-108.	7.0	86
50	An Engineered Device for Indoleacetic Acid Production under Quorum Sensing Signals Enables <i>Cupriavidus pinatubonensis</i> JMP134 To Stimulate Plant Growth. ACS Synthetic Biology, 2018, 7, 1519-1527.	3.8	19
51	Modulating Heterologous Gene Expression with Portable mRNA-Stabilizing 5′-UTR Sequences. ACS Synthetic Biology, 2018, 7, 2177-2188.	3.8	24
52	Biodegradation and Bioremediation: An Introduction. , 2018, , 1-21.		1
53	Re-Factoring Glycolytic Genes for Targeted Engineering of Catabolism in Gram-Negative Bacteria. Methods in Molecular Biology, 2018, 1772, 3-24.	0.9	3
54	Dynamics of <i>Pseudomonas putida</i> biofilms in an upscale experimental framework. Journal of Industrial Microbiology and Biotechnology, 2018, 45, 899-911.	3.0	7

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55	The biofilm matrix polysaccharides cellulose and alginate both protect Pseudomonas putida mt-2 against reactive oxygen species generated under matric stress and copper exposure. Microbiology (United Kingdom), 2018, 164, 883-888.	1.8	33
56	Refactoring the Embden–Meyerhof–Parnas Pathway as a Whole of Portable GlucoBricks for Implantation of Glycolytic Modules in Gram-Negative Bacteria. ACS Synthetic Biology, 2017, 6, 793-805.	3.8	50
57	Deconvolution of Gene Expression Noise into Spatial Dynamics of Transcription Factor–Promoter Interplay. ACS Synthetic Biology, 2017, 6, 1359-1369.	3.8	39
58	Molecular tools and emerging strategies for deep genetic/genomic refactoring of Pseudomonas. Current Opinion in Biotechnology, 2017, 47, 120-132.	6.6	63
59	Bioremediation 3.0: Engineering pollutant-removing bacteria in the times of systemic biology. Biotechnology Advances, 2017, 35, 845-866.	11.7	240
60	Seven microbial bioâ€processes to help the planet. Microbial Biotechnology, 2017, 10, 995-998.	4.2	25
61	CellShape: A userâ€friendly image analysis tool for quantitative visualization of bacterial cell factories inside. Biotechnology Journal, 2017, 12, 1600323.	3.5	15
62	Engineering Gram-Negative Microbial Cell Factories Using Transposon Vectors. Methods in Molecular Biology, 2017, 1498, 273-293.	0.9	23
63	The <scp>RNA</scp> chaperone <scp>Hfq</scp> enables the environmental stress tolerance superâ€phenotype of <scp><i>P</i></scp> <i>seudomonas putida</i> . Environmental Microbiology, 2016, 18, 3309-3326.	3.8	25
64	A Metabolic Widget Adjusts the Phosphoenolpyruvate-Dependent Fructose Influx in Pseudomonas putida. MSystems, 2016, 1, .	3.8	28
65	From dirt to industrial applications: Pseudomonas putida as a Synthetic Biology chassis for hosting harsh biochemical reactions. Current Opinion in Chemical Biology, 2016, 34, 20-29.	6.1	199
66	Nitrogen regulation of the <i>xyl</i> genes of <i>Pseudomonas putida</i> mtâ€2 propagates into a significant effect of nitrate on <i>m</i> â€xylene mineralization in soil. Microbial Biotechnology, 2016, 9, 814-823.	4.2	5
67	Pyridine nucleotide transhydrogenases enable redox balance of <i>Pseudomonas putida</i> during biodegradation of aromatic compounds. Environmental Microbiology, 2016, 18, 3565-3582.	3.8	58
68	The quest for the minimal bacterial genome. Current Opinion in Biotechnology, 2016, 42, 216-224.	6.6	49
69	The Ssr protein (T1E_1405) from <i>Pseudomonas putida</i> DOTâ€T1E enables oligonucleotideâ€based recombineering in platform strain <i>P. putida</i> EM42. Biotechnology Journal, 2016, 11, 1309-1319.	3.5	65
70	The revisited genome of <i>Pseudomonas putida</i> KT2440 enlightens its value as a robust metabolic <i>chassis</i> . Environmental Microbiology, 2016, 18, 3403-3424.	3.8	270
71	Introduction to Systems and Synthetic Biology in Hydrocarbon Microbiology: Applications. Springer Protocols, 2016, , 1-8.	0.3	0
72	Highâ€resolution analysis of the <i>m</i> â€xylene/toluene biodegradation subtranscriptome of <scp><i>P</i></scp> <i>seudomonas putida</i> mtâ€2. Environmental Microbiology, 2016, 18, 3327-3341.	3.8	18

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73	Transcription factor levels enable metabolic diversification of single cells of environmental bacteria. ISME Journal, 2016, 10, 1122-1133.	9.8	13
74	Rationally rewiring the connectivity of the XylR/Pu regulatory node of the m-xylene degradation pathway in Pseudomonas putida. Integrative Biology (United Kingdom), 2016, 8, 571-576.	1.3	0
75	Data on the standardization of a cyclohexanone-responsive expression system for Gram-negative bacteria. Data in Brief, 2016, 6, 738-744.	1.0	17
76	Genetic programming of catalytic Pseudomonas putida biofilms for boosting biodegradation of haloalkanes. Metabolic Engineering, 2016, 33, 109-118.	7.0	103
77	Systems and Synthetic Biology in Hydrocarbon Microbiology: Tools. Springer Protocols, 2015, , 1-7.	0.3	1
78	Plastic waste as a novel substrate for industrial biotechnology. Microbial Biotechnology, 2015, 8, 900-903.	4.2	134
79	Mining Environmental Plasmids for Synthetic Biology Parts and Devices. Microbiology Spectrum, 2015, 3, PLAS-0033-2014.	3.0	18
80	Knock-In-Leave-Behind (KILB): Genetic Grafting of Protease-Cleaving Sequences into Permissive Sites of Proteins with a Tn5-Based Transposition System. Springer Protocols, 2015, , 71-85.	0.3	1
81	Pseudomonas putida mt-2 tolerates reactive oxygen species generated during matric stress by inducing a major oxidative defense response. BMC Microbiology, 2015, 15, 202.	3.3	24
82	The two paralogue <scp><i>phoN</i></scp> (phosphinothricin acetyl transferase) genes of <scp><i>P</i></scp> <i>seudomonas putida</i> encode functionally different proteins. Environmental Microbiology, 2015, 17, 3330-3340.	3.8	6
83	SEVA 2.0: an update of the Standard European Vector Architecture for de-/re-construction of bacterial functionalities. Nucleic Acids Research, 2015, 43, D1183-D1189.	14.5	195
84	Broadening the SEVA Plasmid Repertoire to Facilitate Genomic Editing of Gram-Negative Bacteria. Springer Protocols, 2015, , 9-27.	0.3	9
85	Freeing <scp><i>P</i></scp> <i>seudomonas putida</i> â€ <scp>KT</scp> 2440 of its proviral load strengthens endurance to environmental stresses. Environmental Microbiology, 2015, 17, 76-90.	3.8	62
86	Functional coexistence of twin arsenic resistance systems in <scp><i>P</i></scp> <i>seudomonas putida</i> â€ <scp>KT</scp> 2440. Environmental Microbiology, 2015, 17, 229-238.	3.8	52
87	Genome reduction boosts heterologous gene expression in Pseudomonas putida. Microbial Cell Factories, 2015, 14, 23.	4.0	142
88	Tn7-Based Device for Calibrated Heterologous Gene Expression in <i>Pseudomonas putida</i> . ACS Synthetic Biology, 2015, 4, 1341-1351.	3.8	169
89	Phenotypic knockouts of selected metabolic pathways by targeting enzymes with camel-derived nanobodies (VHHs). Metabolic Engineering, 2015, 30, 40-48.	7.0	8
90	Pseudomonas putida KT2440 Strain Metabolizes Glucose through a Cycle Formed by Enzymes of the Entner-Doudoroff, Embden-Meyerhof-Parnas, and Pentose Phosphate Pathways. Journal of Biological Chemistry, 2015, 290, 25920-25932.	3.4	269

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91	The Glycerol-Dependent Metabolic Persistence of Pseudomonas putida KT2440 Reflects the Regulatory Logic of the GlpR Repressor. MBio, 2015, 6, .	4.1	62
92	Widening functional boundaries of the $I_f$ (sup>54 /sup>promoter Pu of Pseudomonas putida by defeating extant physiological constraints. Molecular BioSystems, 2015, 11, 734-742.	2.9	4
93	The differential response of the <scp><i>P</i></scp> <i>ben</i> promoter of <scp><i>P</i></scp> <i>seudomonas putida</i> â€ <scp>mt</scp> â€2 to <scp>BenR</scp> and <scp>XylSprevents metabolic conflicts in <scp><i>m</i></scp><i>â€</i>xylene biodegradation. Environmental Microbiology. 2015. 17. 64-75.</scp>	0> 3.8	29
94	Chassis organism from <i>Corynebacterium glutamicum</i> : The way towards biotechnological domestication of Corynebacteria. Biotechnology Journal, 2015, 10, 244-245.	3.5	11
95	Confidence, tolerance, and allowance in biological engineering: The nuts and bolts of living things. BioEssays, 2015, 37, 95-102.	2.5	22
96	Metabolic and regulatory rearrangements underlying glycerol metabolism in <i><scp>P</scp>seudomonas putida</i> â€ <scp>KT</scp> 2440. Environmental Microbiology, 2014, 16, 239-254.	3.8	91
97	Chemical reactivity drives spatiotemporal organisation of bacterial metabolism. FEMS Microbiology Reviews, 2014, 39, n/a-n/a.	8.6	67
98	Pseudomonas 2.0: genetic upgrading of P. putida KT2440 as an enhanced host for heterologous gene expression. Microbial Cell Factories, 2014, 13, 159.	4.0	199
99	Robustness of Pseudomonas putida KT2440 as a host for ethanol biosynthesis. New Biotechnology, 2014, 31, 562-571.	4.4	62
100	Engineering Multicellular Logic in Bacteria with Metabolic Wires. ACS Synthetic Biology, 2014, 3, 204-209.	3.8	30
101	From the <i>selfish gene</i> to <i>selfish metabolism</i> : Revisiting the central dogma. BioEssays, 2014, 36, 226-235.	2.5	60
102	The private life of environmental bacteria: pollutant biodegradation at the single cell level. Environmental Microbiology, 2014, 16, 628-642.	3.8	63
103	Biotechnological domestication of pseudomonads using synthetic biology. Nature Reviews Microbiology, 2014, 12, 368-379.	28.6	332
104	From the phosphoenolpyruvate phosphotransferase system to selfish metabolism: a story retraced in <i>Pseudomonas putida</i> . FEMS Microbiology Letters, 2014, 356, 144-153.	1.8	26
105	The metabolic cost of flagellar motion in <scp><i>P</i></scp> <i>seudomonas putida</i> â€ <scp>KT</scp> 2440. Environmental Microbiology, 2014, 16, 291-303.	3.8	132
106	Volatilization of Arsenic from Polluted Soil by <i>Pseudomonas putida</i> Engineered for Expression of the <i>arsM</i> Arsenic(III) S-Adenosine Methyltransferase Gene. Environmental Science & Technology, 2014, 48, 10337-10344.	10.0	106
107	Fructose 1â€phosphate is the one and only physiological effector of the Cra (FruR) regulator of <i>Pseudomonas putida</i> . FEBS Open Bio, 2014, 4, 377-386.	2.3	28
108	New Transposon Tools Tailored for Metabolic Engineering of Gram-Negative Microbial Cell Factories. Frontiers in Bioengineering and Biotechnology, 2014, 2, 46.	4.1	85

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109	The Standard European Vector Architecture (SEVA) Plasmid Toolkit. Methods in Molecular Biology, 2014, 1149, 469-478.	0.9	28
110	Chromosomal Integration of Transcriptional Fusions. Methods in Molecular Biology, 2014, 1149, 479-489.	0.9	7
111	Promoter Fusions with Optical Outputs in Individual Cells and in Populations. Methods in Molecular Biology, 2014, 1149, 579-590.	0.9	1
112	The IHF regulon of exponentially growing <i>Pseudomonas putida</i> cells. Environmental Microbiology, 2013, 15, 49-63.	3.8	14
113	Cra regulates the crossâ€ŧalk between the two branches of the phosphoenolpyruvate : phosphotransferase system of <i>Pseudomonas putida</i> . Environmental Microbiology, 2013, 15, 121-132.	3.8	18
114	Accumulation of inorganic polyphosphate enables stress endurance and catalytic vigour in Pseudomonas putida KT2440. Microbial Cell Factories, 2013, 12, 50.	4.0	77
115	Why are chlorinated pollutants so difficult to degrade aerobically? Redox stress limits 1,3-dichloprop-1-ene metabolism by <i>Pseudomonas pavonaceae</i> . Philosophical Transactions of the Royal Society B: Biological Sciences, 2013, 368, 20120377.	4.0	53
116	Expanding the Boolean Logic of the Prokaryotic Transcription Factor XylR by Functionalization of Permissive Sites with a Protease-Target Sequence. ACS Synthetic Biology, 2013, 2, 594-603.	3.8	16
117	NanoPad: An integrated platform for bacterial production of camel nanobodies aimed at detecting environmental biomarkers. Proteomics, 2013, 13, 2766-2775.	2.2	7
118	The <scp>E</scp> ntner– <scp>D</scp> oudoroff pathway empowers <i><scp>P</scp>seudomonas putida</i> â€ <scp>KT</scp> 2440 with a high tolerance to oxidative stress. Environmental Microbiology, 2013, 15, 1772-1785.	3.8	195
119	Implantation of unmarked regulatory and metabolic modules in Gram-negative bacteria with specialised mini-transposon delivery vectors. Journal of Biotechnology, 2013, 163, 143-154.	3.8	51
120	Engineering an anaerobic metabolic regime in Pseudomonas putida KT2440 for the anoxic biodegradation of 1,3-dichloroprop-1-ene. Metabolic Engineering, 2013, 15, 98-112.	7.0	93
121	Endogenous Stress Caused by Faulty Oxidation Reactions Fosters Evolution of 2,4-Dinitrotoluene-Degrading Bacteria. PLoS Genetics, 2013, 9, e1003764.	3.5	74
122	Towards Functional Orthogonalisation of Protein Complexes: Individualisation of GroEL Monomers Leads to Distinct Quasihomogeneous Single Rings. ChemBioChem, 2013, 14, 2310-2321.	2.6	10
123	Vestigialization of arsenic resistance phenotypes/genotypes inChromobacterium violaceumstrains thriving in pristine Brazilian sites. Biocatalysis and Biotransformation, 2013, 31, 281-291.	2.0	2
124	Transcriptomic fingerprinting of <i><scp>P</scp>seudomonas putida</i> under alternative physiological regimes. Environmental Microbiology Reports, 2013, 5, 883-891.	2.4	75
125	Engineering the Soil Bacterium Pseudomonas putida for Arsenic Methylation. Applied and Environmental Microbiology, 2013, 79, 4493-4495.	3.1	85
126	The Standard European Vector Architecture (SEVA): a coherent platform for the analysis and deployment of complex prokaryotic phenotypes. Nucleic Acids Research, 2013, 41, D666-D675.	14.5	556

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127	Increasing Signal Specificity of the TOL Network of Pseudomonas putida mt-2 by Rewiring the Connectivity of the Master Regulator XylR. PLoS Genetics, 2012, 8, e1002963.	3.5	26
128	Regulatory Tasks of the Phosphoenolpyruvate-Phosphotransferase System of Pseudomonas putida in Central Carbon Metabolism. MBio, 2012, 3, .	4.1	78
129	A GFP-lacZ Bicistronic Reporter System for Promoter Analysis in Environmental Gram-Negative Bacteria. PLoS ONE, 2012, 7, e34675.	2.5	19
130	Stochasticity of TOL plasmid catabolic promoters sets a bimodal expression regime in <i>Pseudomonas putida</i> mtâ€2 exposed to <i>mâ€</i> xylene. Molecular Microbiology, 2012, 86, 199-211.	2.5	20
131	Transposon-Based and Plasmid-Based Genetic Tools for Editing Genomes of Gram-Negative Bacteria. Methods in Molecular Biology, 2012, 813, 267-283.	0.9	92
132	Broadening the signal specificity of prokaryotic promoters by modifying cis-regulatory elements associated with a single transcription factor. Molecular BioSystems, 2012, 8, 1950.	2.9	30
133	Engineering Whole-Cell Biosensors with No Antibiotic Markers for Monitoring Aromatic Compounds in the Environment. Methods in Molecular Biology, 2012, 834, 261-281.	0.9	19
134	Quantitative, Non-Disruptive Monitoring of Transcription in Single Cells with a Broad-Host Range GFP-luxCDABE Dual Reporter System. PLoS ONE, 2012, 7, e52000.	2.5	22
135	Synthetic constructs in/for the environment: Managing the interplay between natural and engineered Biology. FEBS Letters, 2012, 586, 2199-2206.	2.8	78
136	The Crp regulator of <i>Pseudomonas putida</i> : evidence of an unusually high affinity for its physiological effector, cAMP. Environmental Microbiology, 2012, 14, 702-713.	3.8	14
137	Random and cyclical deletion of large DNA segments in the genome of <i>Pseudomonas putida</i> . Environmental Microbiology, 2012, 14, 1444-1453.	3.8	56
138	A composite feed-forward loop I4-FFL involving IHF and Crc stabilizes expression of the XylR regulator of Pseudomonas putida mt-2 from growth phase perturbations. Molecular BioSystems, 2011, 7, 2982.	2.9	21
139	Implementing an OR–NOT (ORN) logic gate with components of the SOS regulatory network of Escherichia coli. Molecular BioSystems, 2011, 7, 2389.	2.9	18
140	The interplay of the EllANtr component of the nitrogen-related phosphotransferase system (PTSNtr) of Pseudomonas putida with pyruvate dehydrogenase. Biochimica Et Biophysica Acta - General Subjects, 2011, 1810, 995-1005.	2.4	32
141	Regulatory exaptation of the catabolite repression protein (Crp)–cAMP system in <i>Pseudomonas putida</i> . Environmental Microbiology, 2011, 13, 324-339.	3.8	34
142	Monitoring biodegradative enzymes with nanobodies raised in <i>Camelus dromedarius</i> with mixtures of catabolic proteins. Environmental Microbiology, 2011, 13, 960-974.	3.8	21
143	The <i>logicome</i> of environmental bacteria: merging catabolic and regulatory events with Boolean formalisms. Environmental Microbiology, 2011, 13, 2389-2402.	3.8	36
144	Engineering multiple genomic deletions in Gramâ€negative bacteria: analysis of the multiâ€resistant antibiotic profile of <i>Pseudomonas putida</i> KT2440. Environmental Microbiology, 2011, 13, 2702-2716.	3.8	329

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145	Cooperative amino acid changes shift the response of the σ <sup>54</sup> â€dependent regulator XylR from natural <i>mâ€</i> xylene towards xenobiotic 2,4â€dinitrotoluene. Molecular Microbiology, 2011, 79, 1248-1259.	2.5	26
146	Association of dnt genes of Burkholderia sp. DNT with the substrate-blind regulator DntR draws the evolutionary itinerary of 2,4-dinitrotoluene biodegradation. Molecular Microbiology, 2011, 82, 287-299.	2.5	29
147	Functional analysis of the integration host factor site of the Ïf <sup>54</sup> <i>Pu</i> promoter of <i>Pseudomonas putida</i> by <i>in vivo</i> UV imprinting. Molecular Microbiology, 2011, 82, 591-601.	2.5	7
148	pBAM1: an all-synthetic genetic tool for analysis and construction of complex bacterial phenotypes. BMC Microbiology, 2011, 11, 38.	3.3	142
149	In situ detection of aromatic compounds with biosensor Pseudomonas putida cells preserved and delivered to soil in water-soluble gelatin capsules. Analytical and Bioanalytical Chemistry, 2011, 400, 1093-1104.	3.7	34
150	The logic layout of the TOL network of Pseudomonas putida pWW0 plasmid stems from a metabolic amplifier motif (MAM) that optimizes biodegradation of m-xylene. BMC Systems Biology, 2011, 5, 191.	3.0	32
151	Beware of metaphors: Chasses and orthogonality in synthetic biology. Bioengineered Bugs, 2011, 2, 3-7.	1.7	64
152	Environmental biosafety in the age of Synthetic Biology: Do we really need a radical new approach?. BioEssays, 2010, 32, 926-931.	2.5	37
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